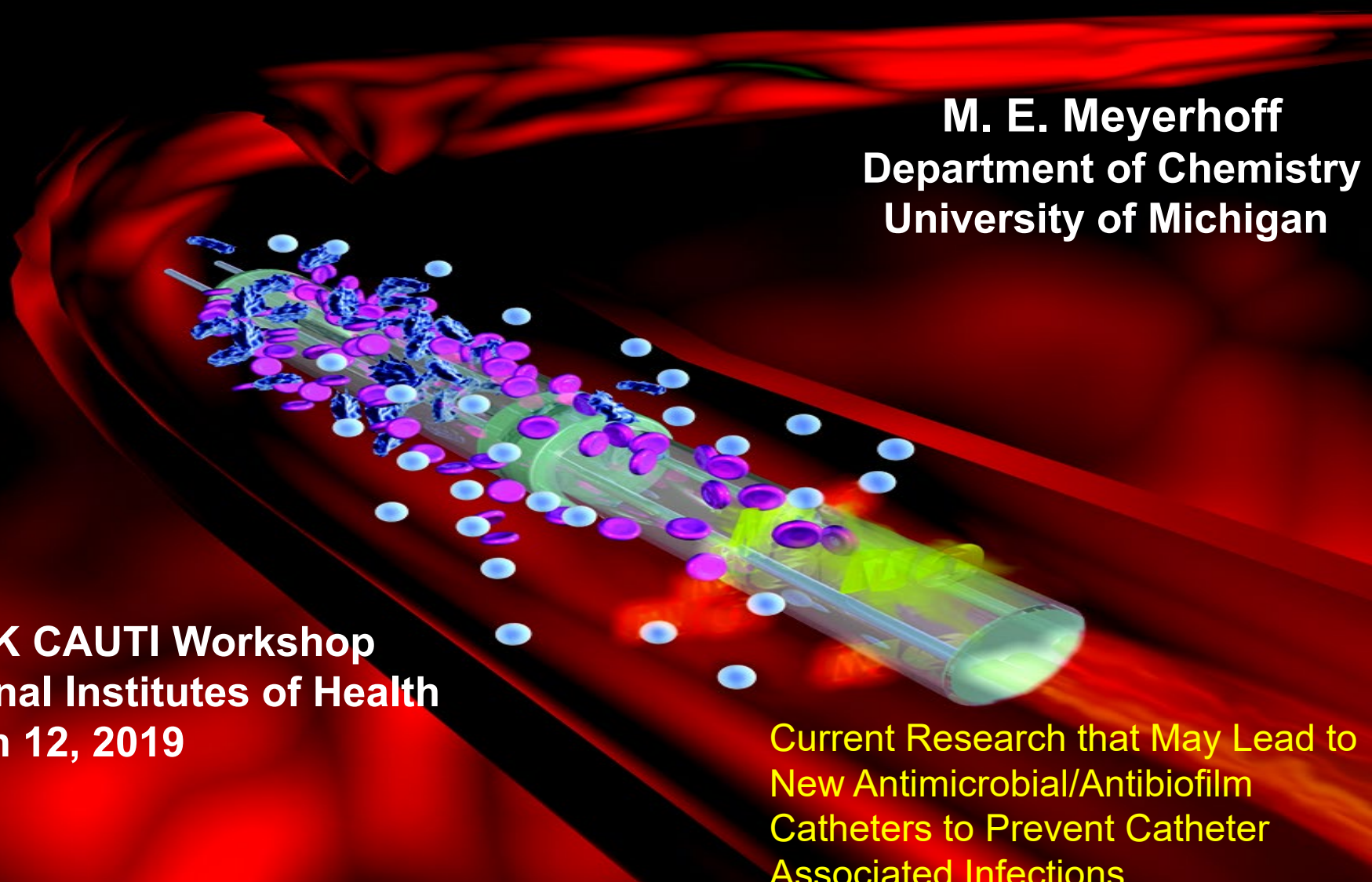


Nitric Oxide (NO) Releasing Urinary Catheters to Reduce the Risk of CAUTIs

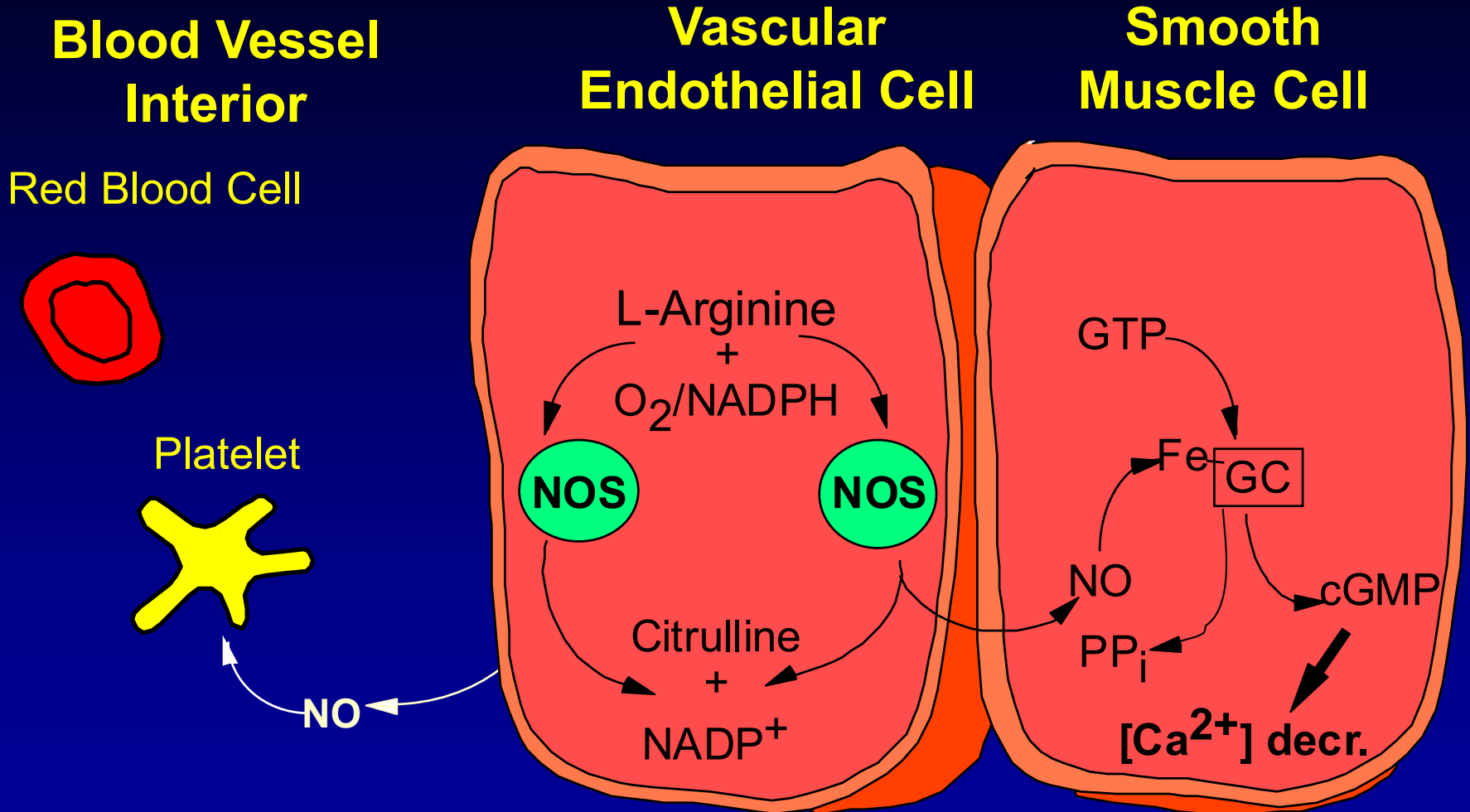
M. E. Meyerhoff
Department of Chemistry
University of Michigan



NIDDK CAUTI Workshop
National Institutes of Health
March 12, 2019

Current Research that May Lead to
New Antimicrobial/Antibiofilm
Catheters to Prevent Catheter
Associated Infections

NO Production: Endothelial Cells (EDRF)



surface flux:

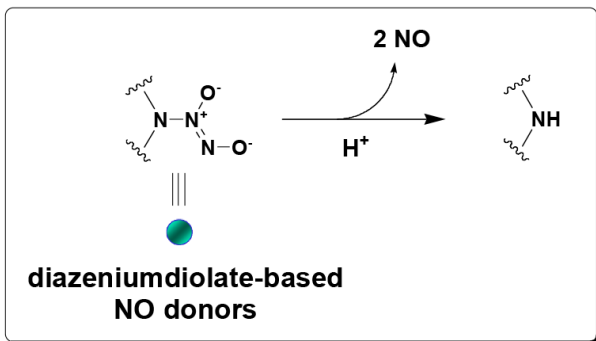
$1 \times 10^{-10} \text{ mol/cm}^2\text{min}$

NOS : Nitric Oxide Synthase

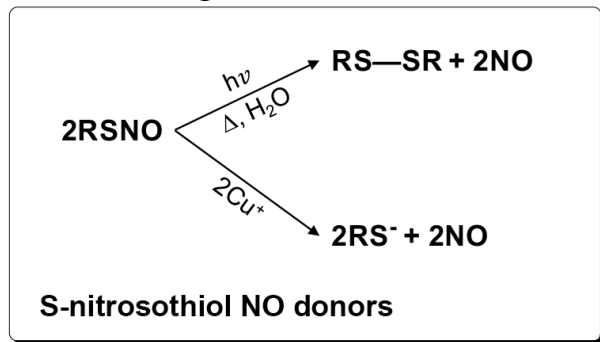
Nitric Oxide Release Chemistries Examined

Chemistry

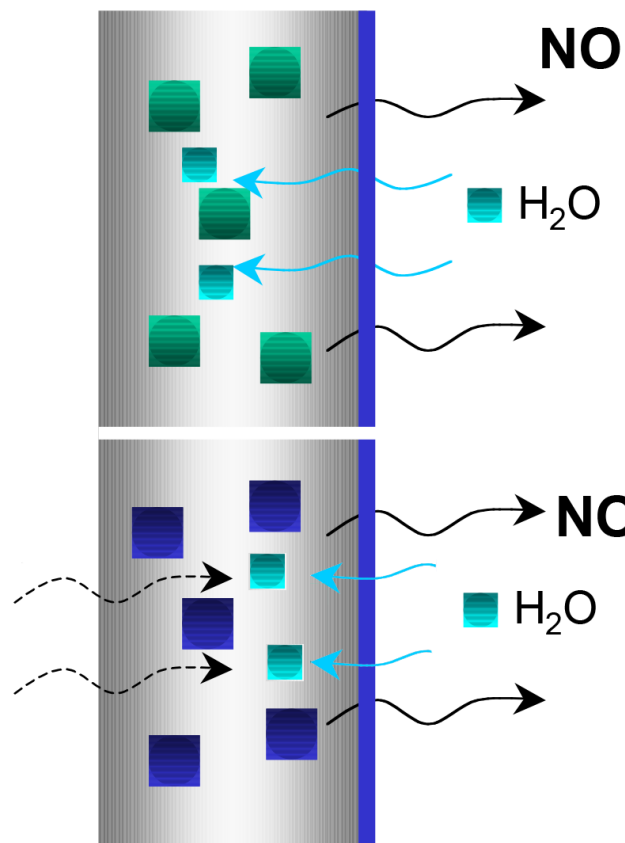
I. Proton-mediated NO release



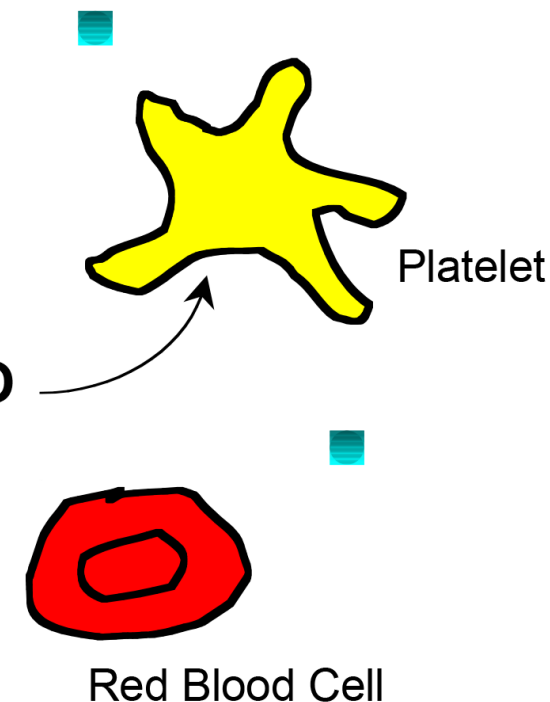
II. Thermal/Light/Water NO release



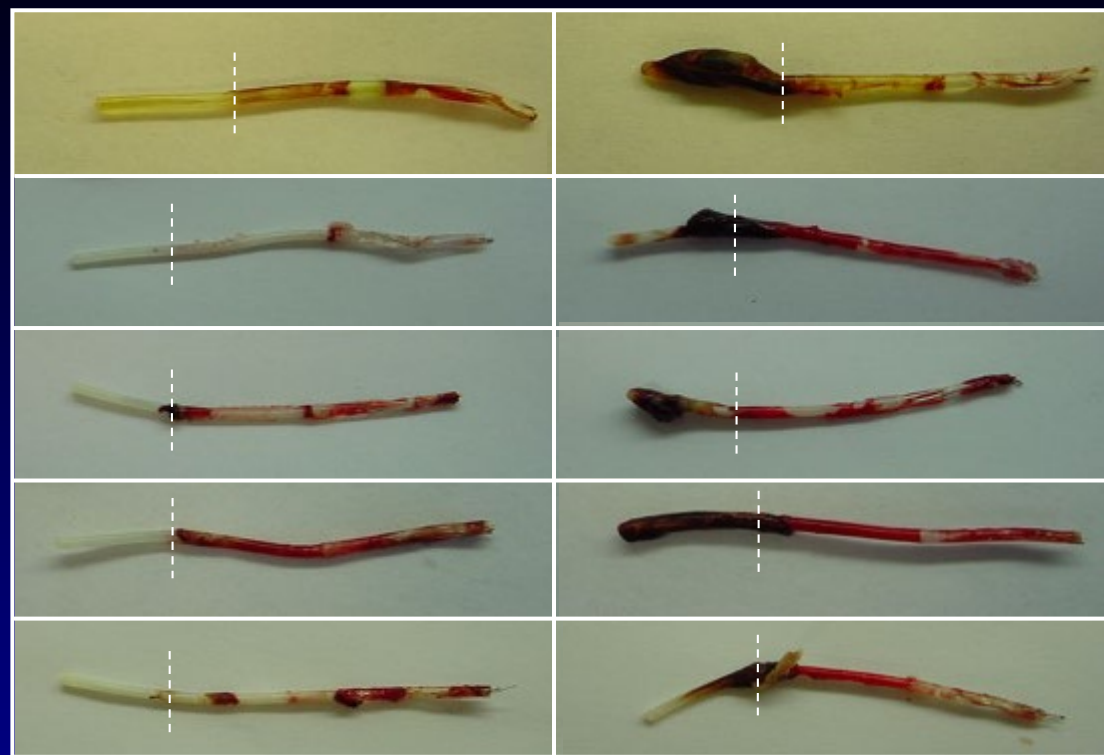
Polymer phase



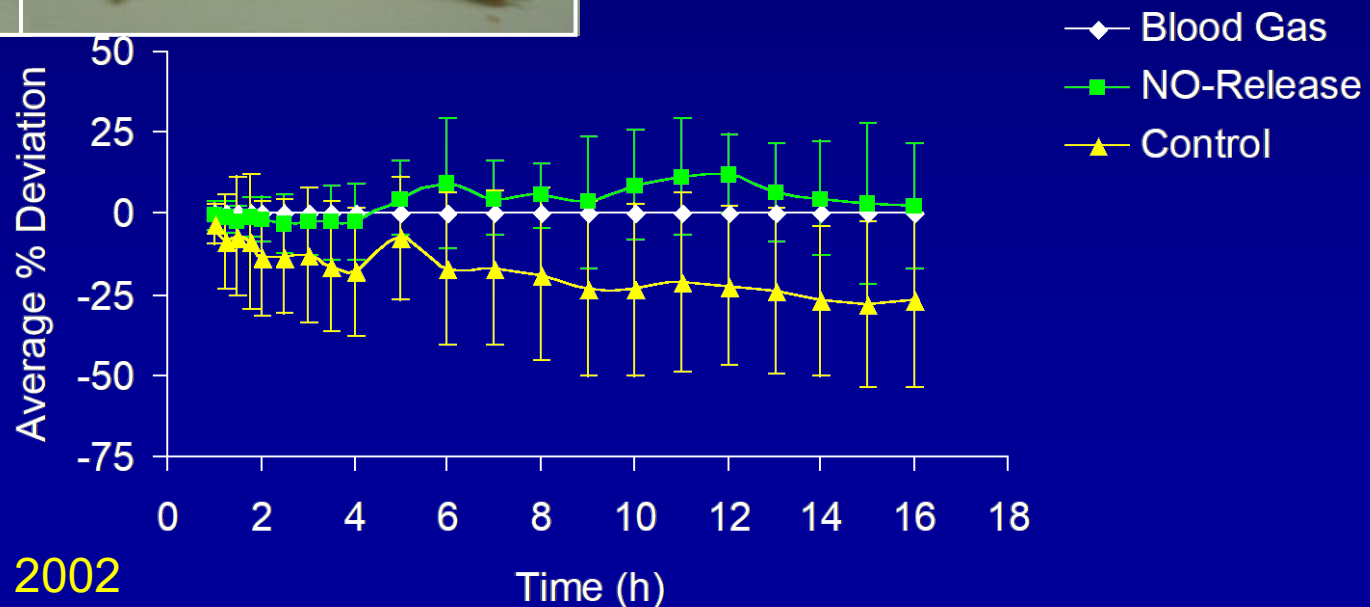
Blood phase



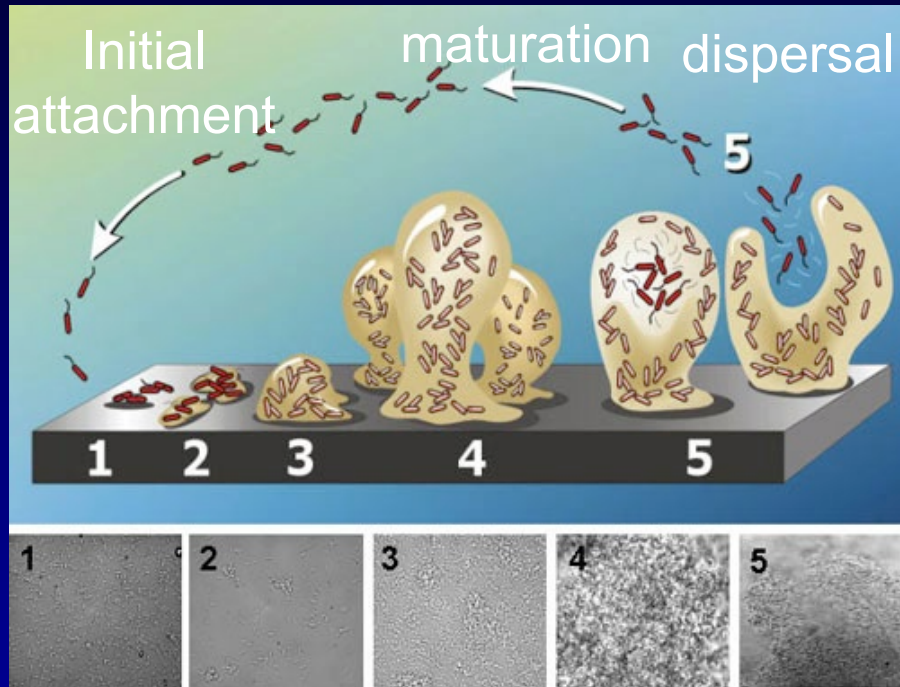
a) With NO-Release b) Without NO-Release



Oxygen Sensors After
Removal From
Porcine Arteries (16-18 h)



Bacterial Biofilms on Medical Devices: Can't be Treated Effectively with Antibiotics

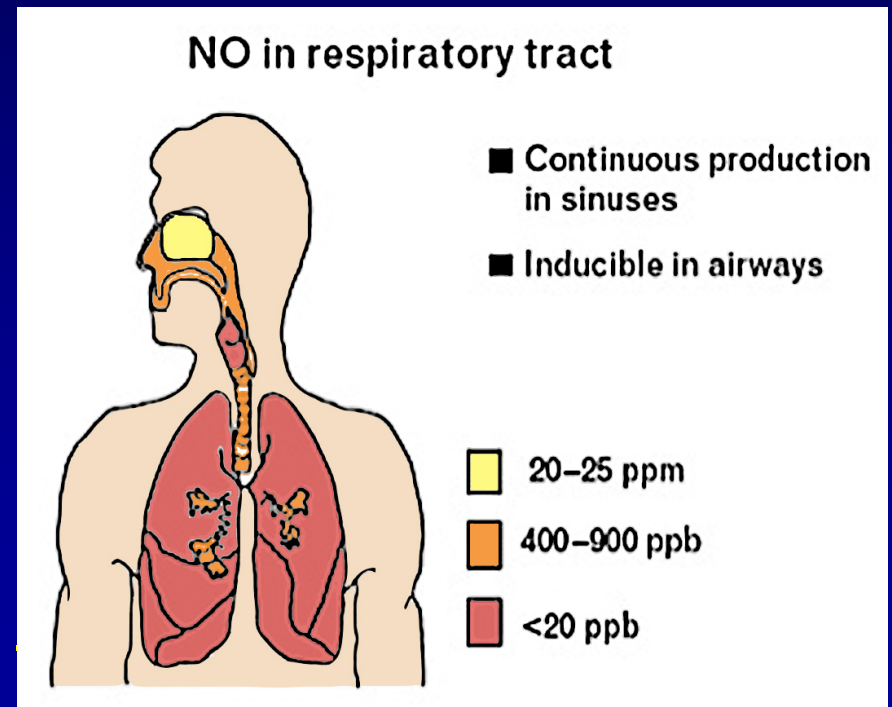


Biofilm Development Stages

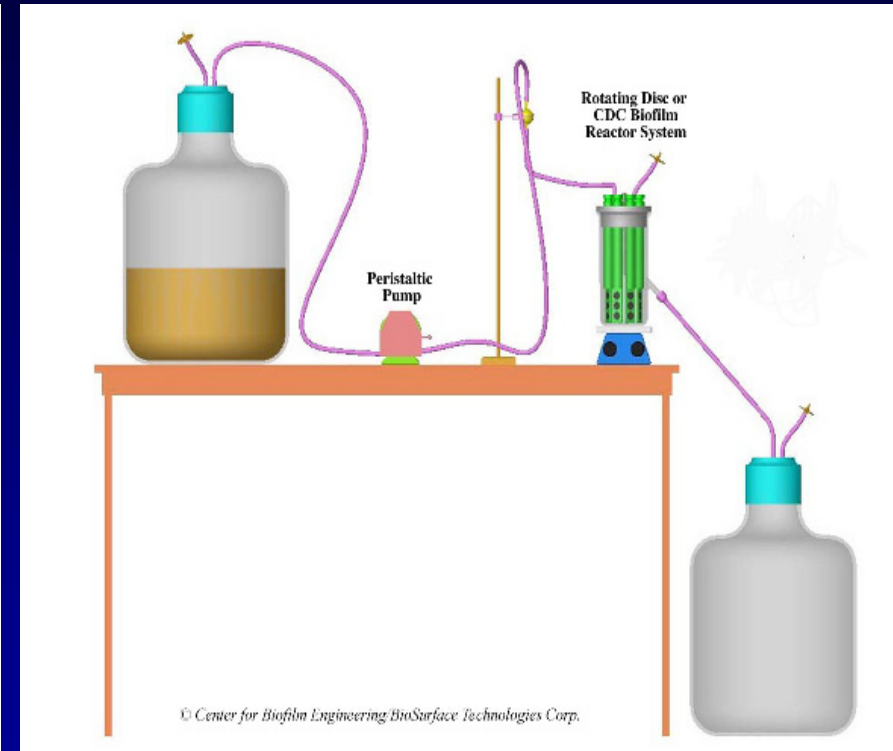
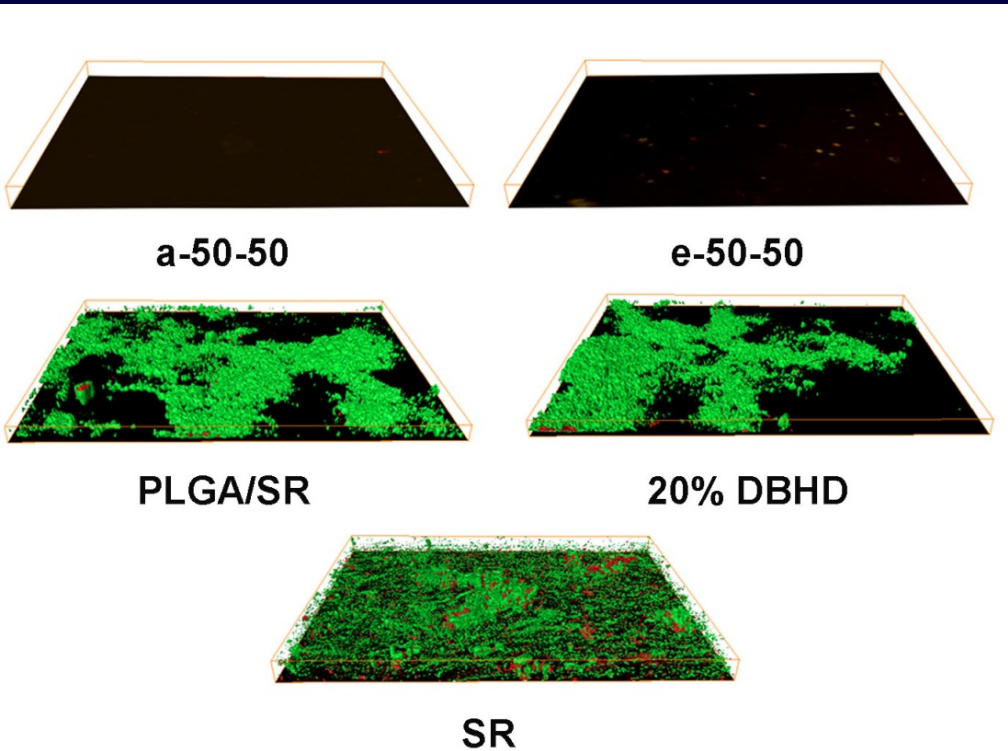
**Macrophages produce high NO levels:
This is how we fight infection!!**

Nitric Oxide

**Natural and potent
antimicrobial and
biofilm dispersal agent**



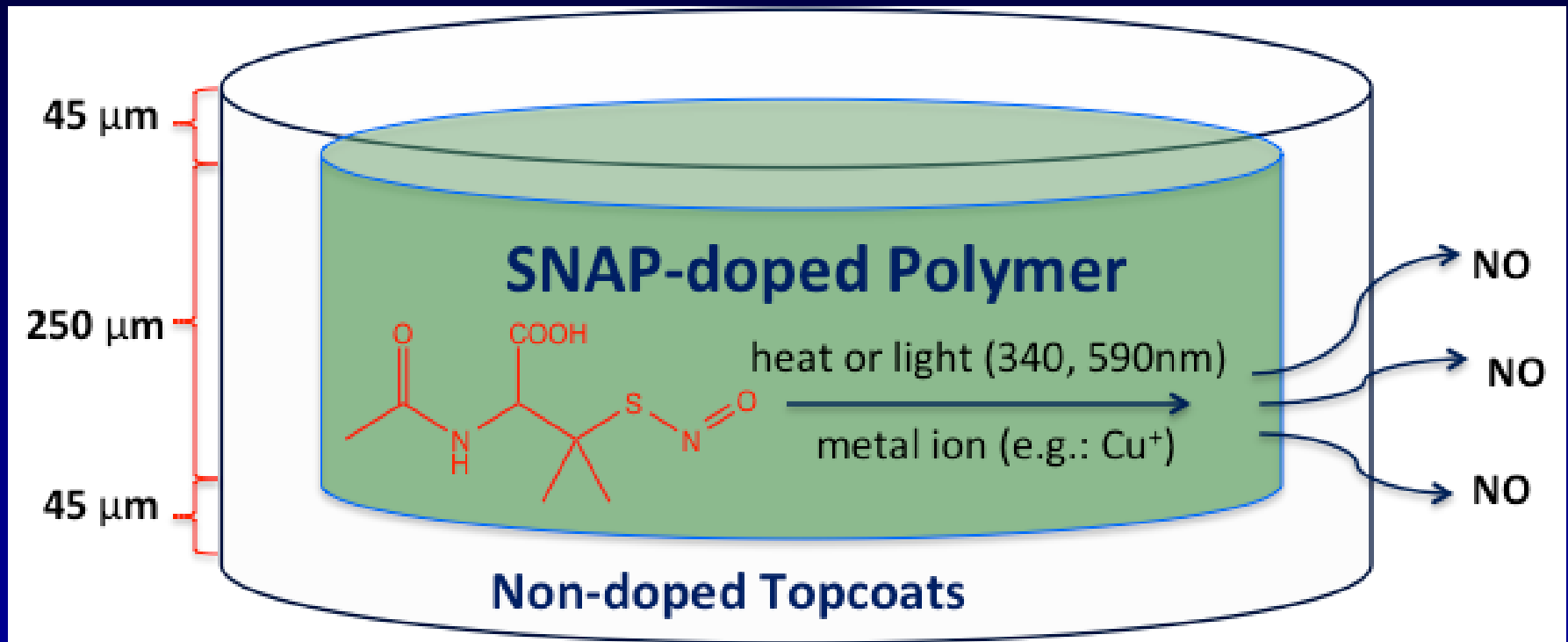
Chemical Release of NO Significantly Reduces Microbial Biofilm (*S. aureus*) Formation over 7 d (DBHD/NONO in SR Films-w/PLGA Additive)



S. aureus biofilm formation on surfaces of NO releasing (top) and control silicone rubber films (lower three films are controls)

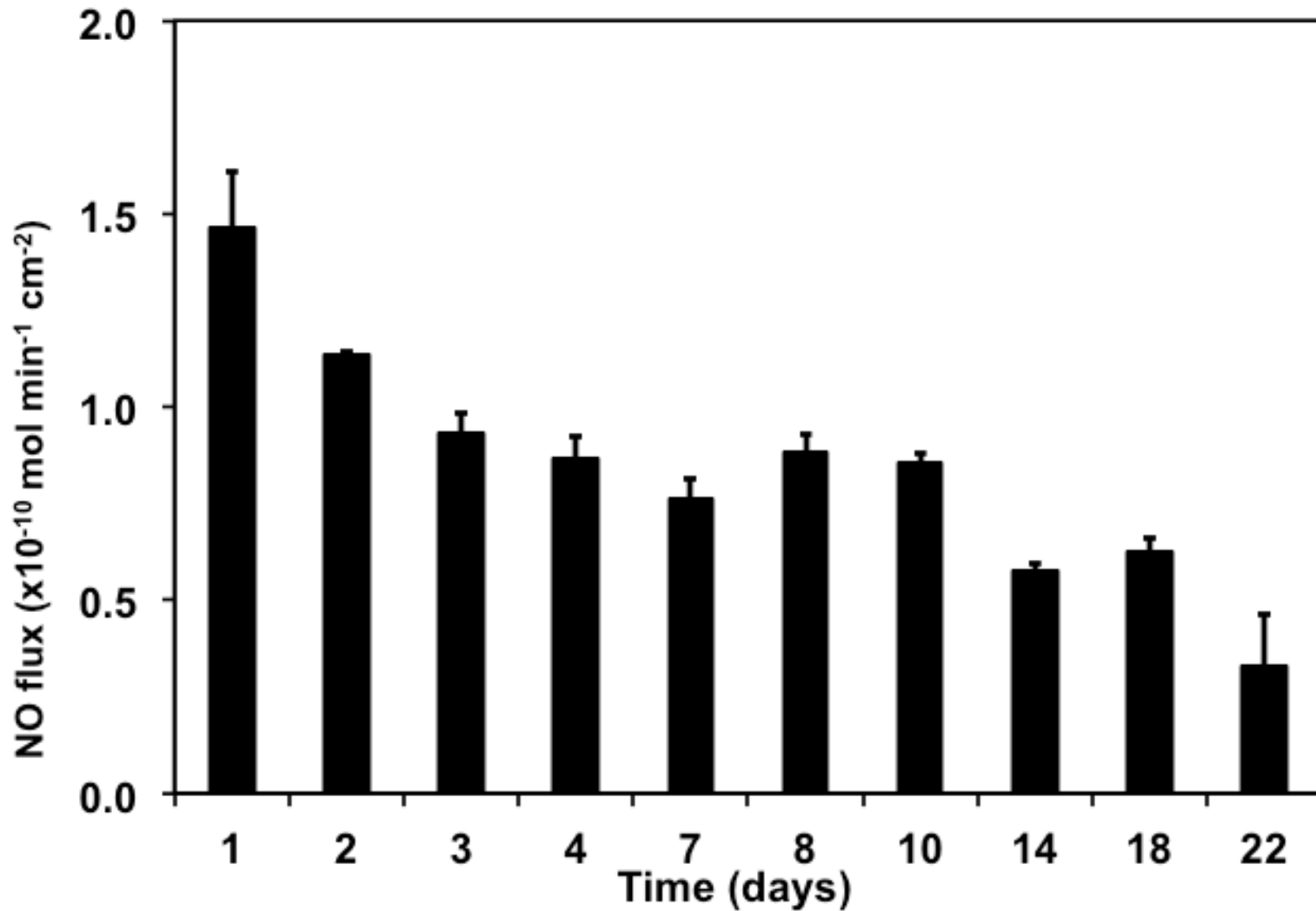
CDC Biofilm Reactor

s-S-Nitroso-N-Acetylpenicillamine (SNAP) Doped Polymers



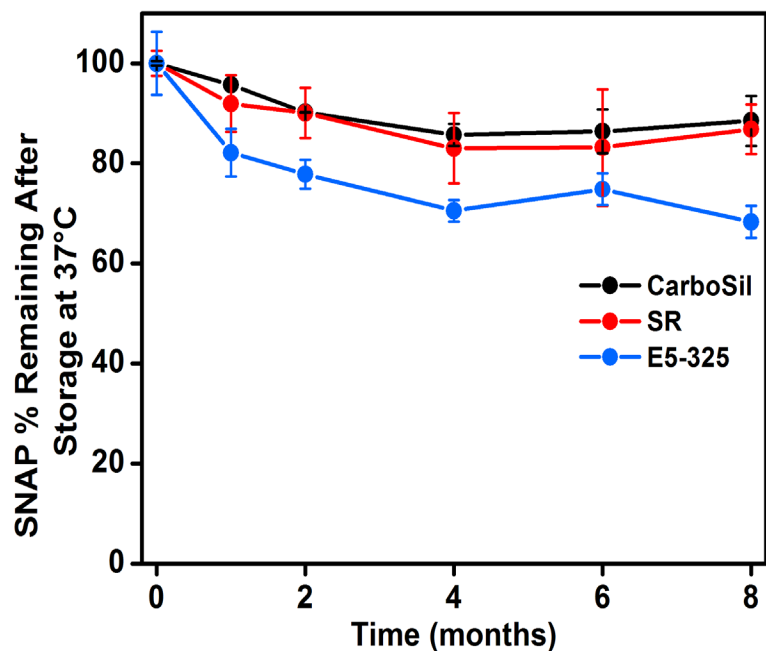
10 wt% SNAP-Doped CarboSil Film

NO Release at 37 °C (kept in PBS buffer)

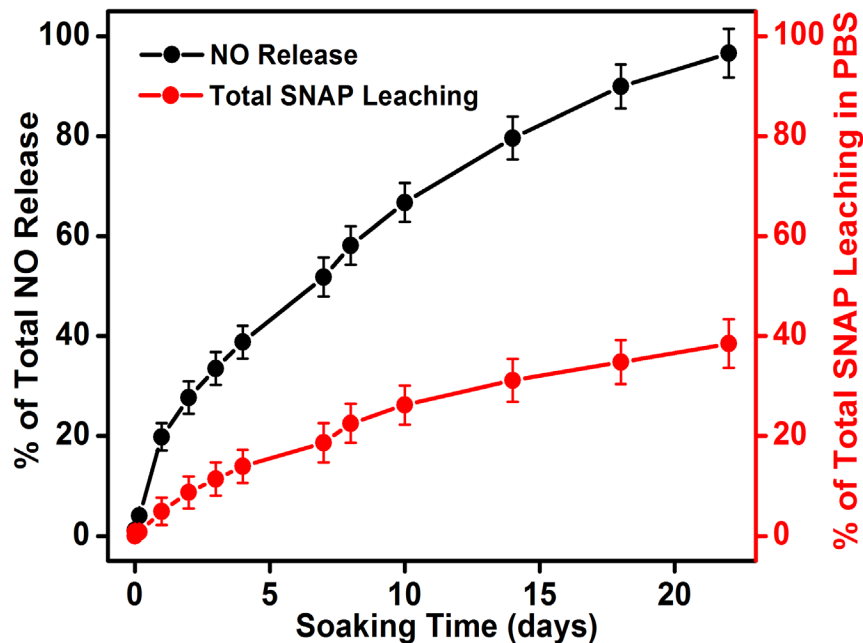


SNAP-Doped CarboSil Composite Material (Long-Term Behavior)

Shelf-Life Stability

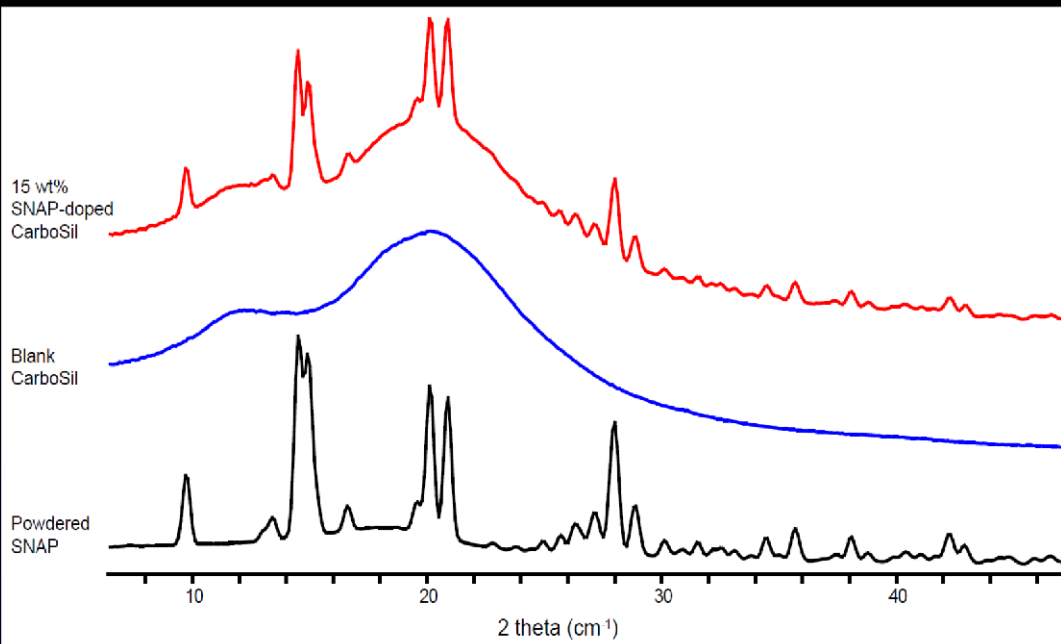


SNAP/NAP/(NAP)₂ Leaching

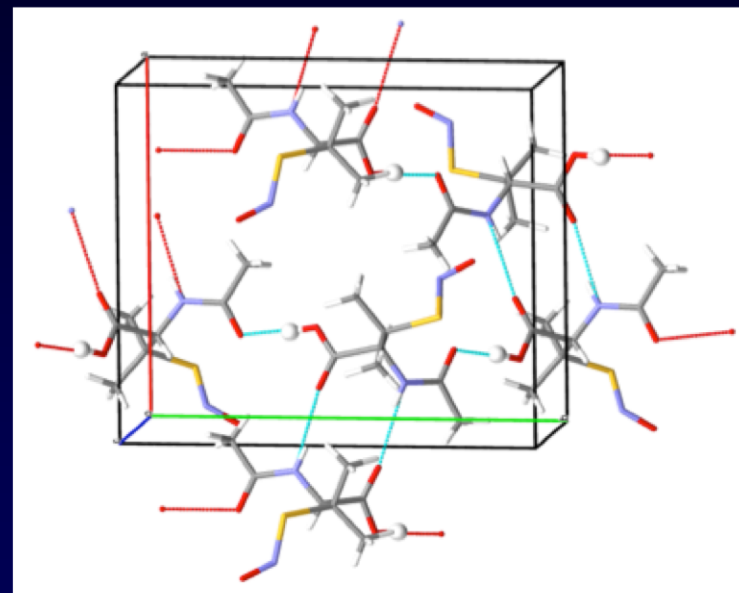
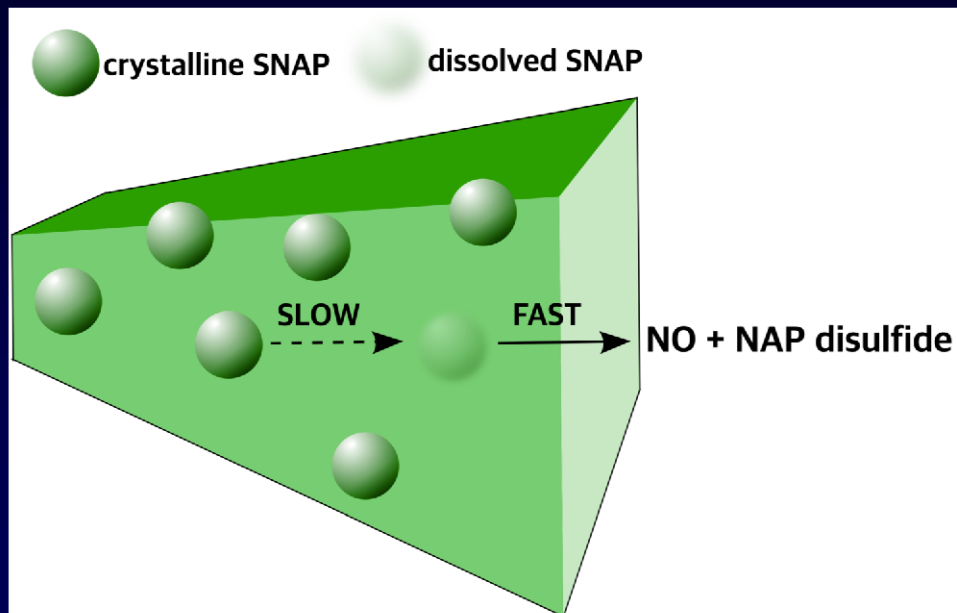
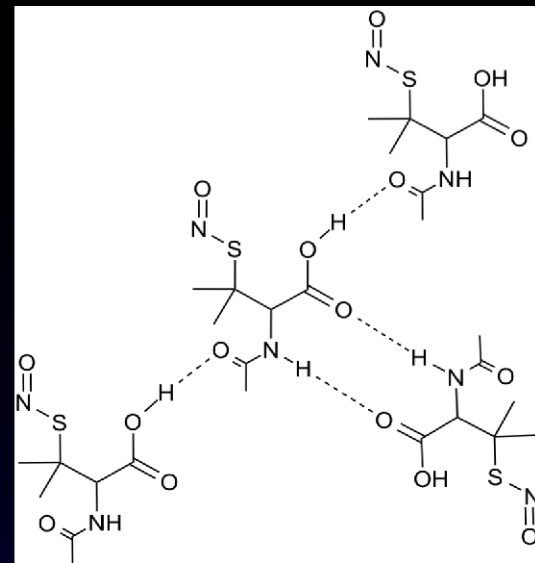


stored dry in dark at 37 °C

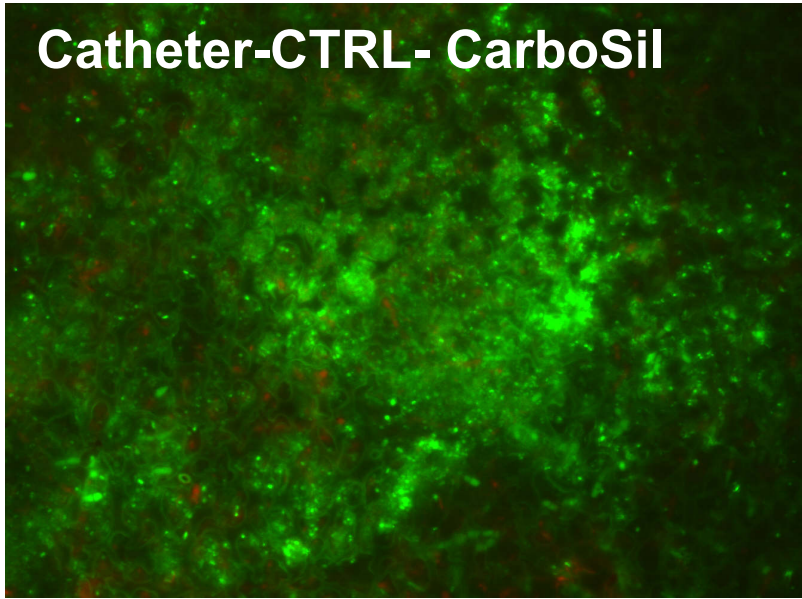
-stored in PBS (pH 7.4) at 37 °C
-soaking solutions changed daily
-SNAP/NAP/(NAP)₂ determined by LCMS



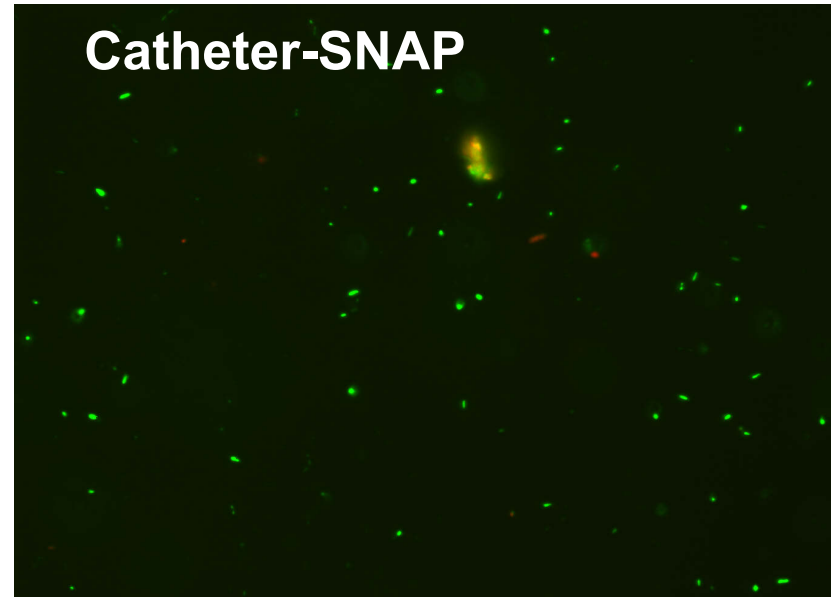
PXRD patterns of SNAP powder, blank CarboSil and 15 wt% SNAP-doped CarboSil film samples.



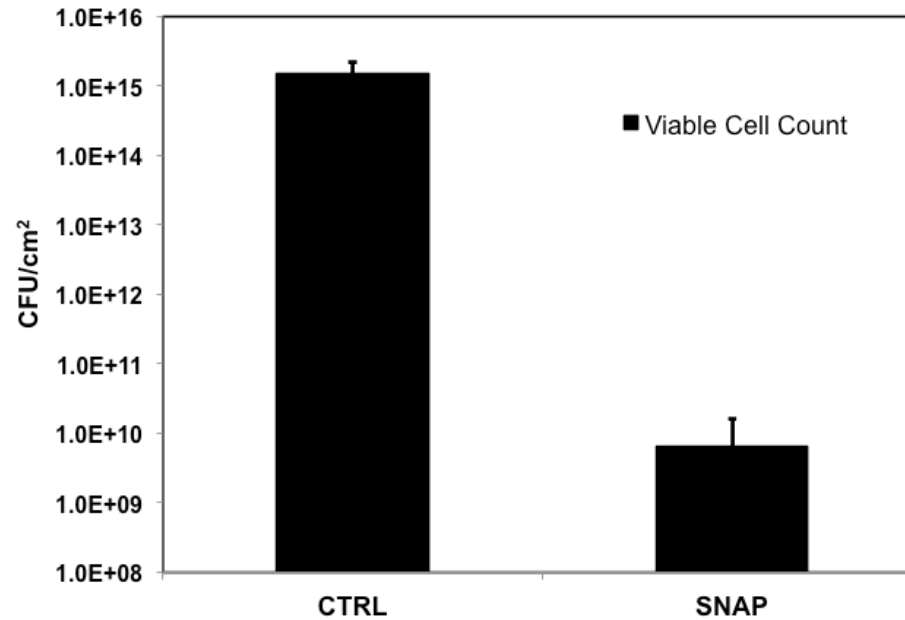
Catheter-CTRL- CarboSil



Catheter-SNAP

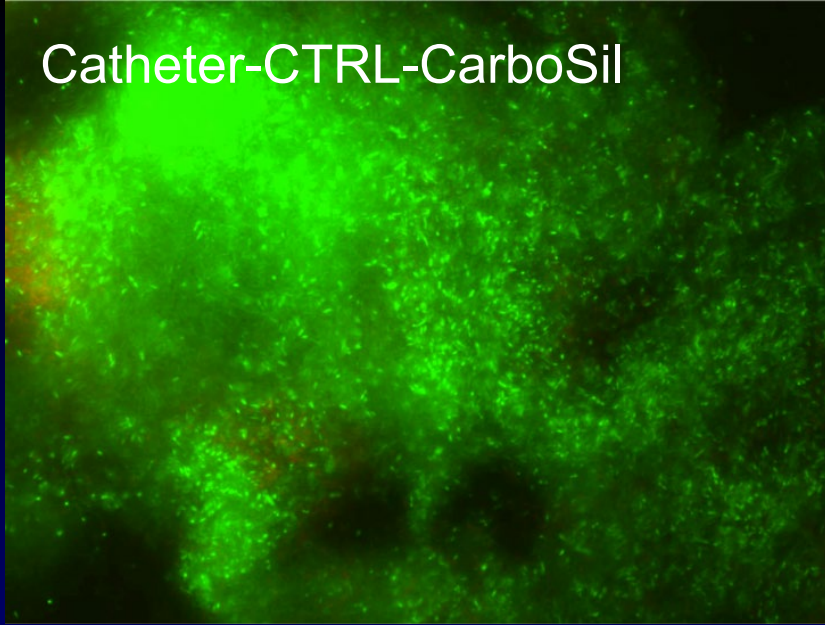


7 Days *S. aureus* Biofilm Plate Count (n=4)

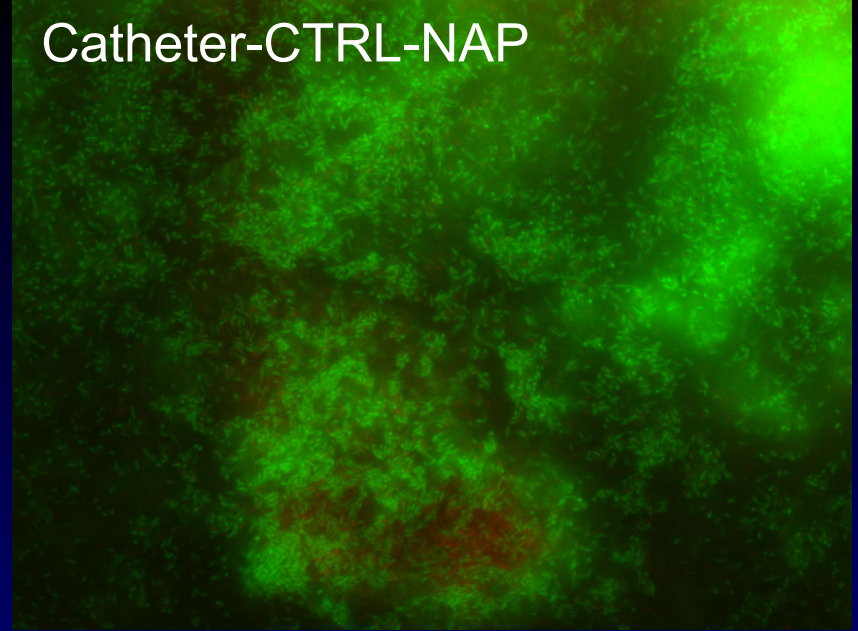


drip-flow
bioreactor

Catheter-CTRL-CarboSil

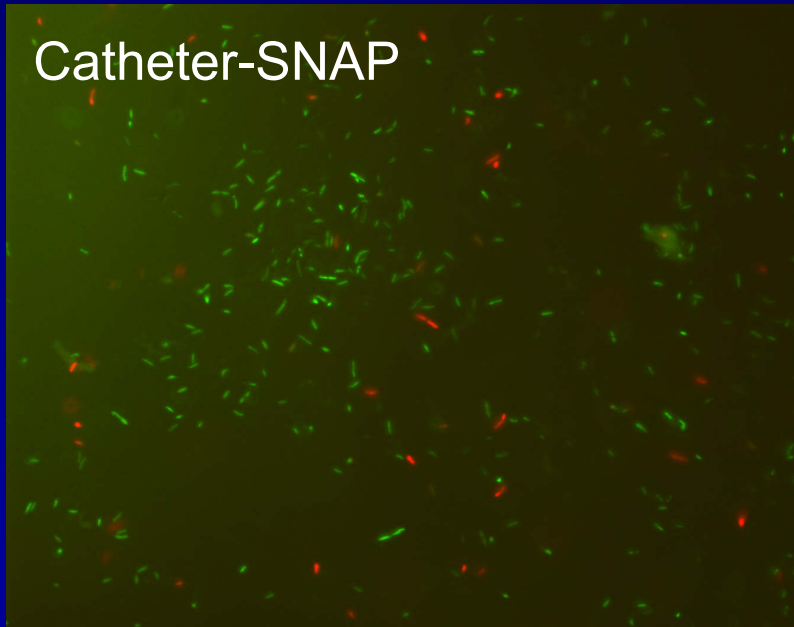


Catheter-CTRL-NAP

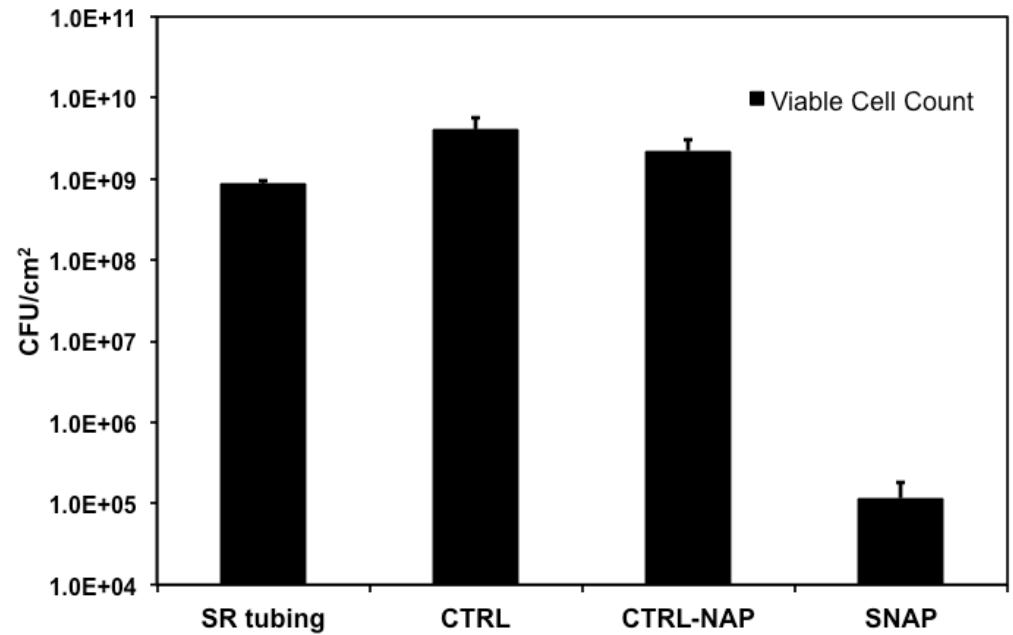


CDC Bioreactor

Catheter-SNAP



7 Days *P.aeruginosa* Biofilm Plate Count (n=4)



S-Nitroso-N-acetylpenicillamine (SNAP) Impregnated Silicone Foley Catheters: A Potential Biomaterial/Device To Prevent Catheter-Associated Urinary Tract Infections

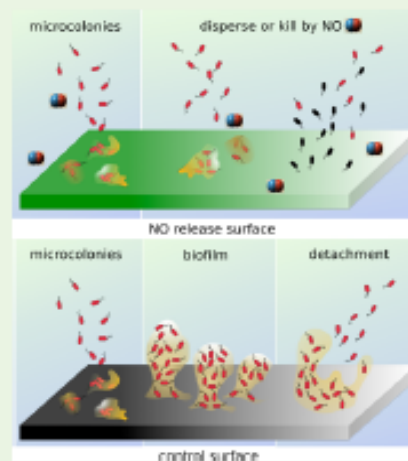
Alessandro Colletta,[†] Jianfeng Wu,[‡] Yaqi Wo,[†] Michael Kappler,[§] Hao Chen,[§] Chuanwu Xi,^{*,‡} and Mark E. Meyerhoff^{*,†}

[†]Department of Chemistry and [‡]Department of Environmental Health Sciences, University of Michigan, Ann Arbor, Michigan 48109, United States

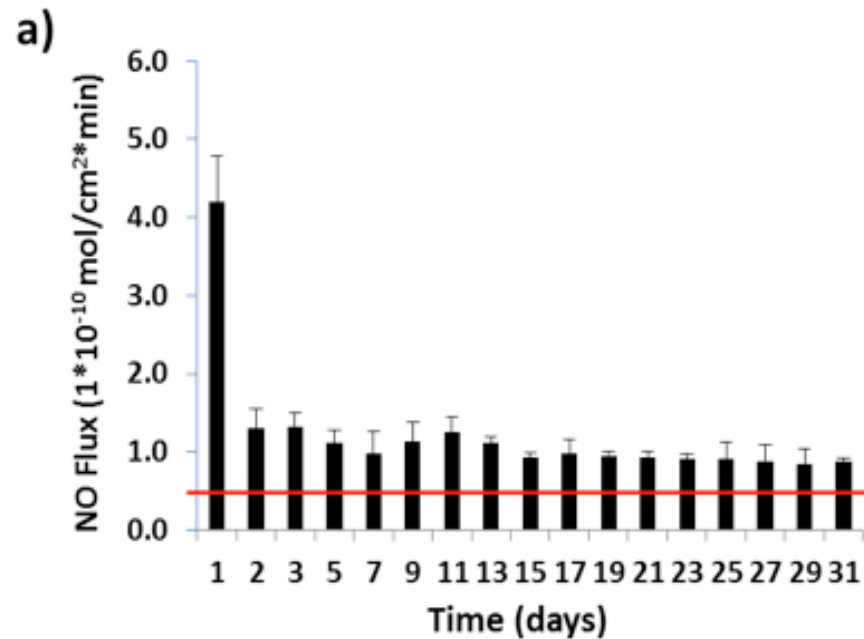
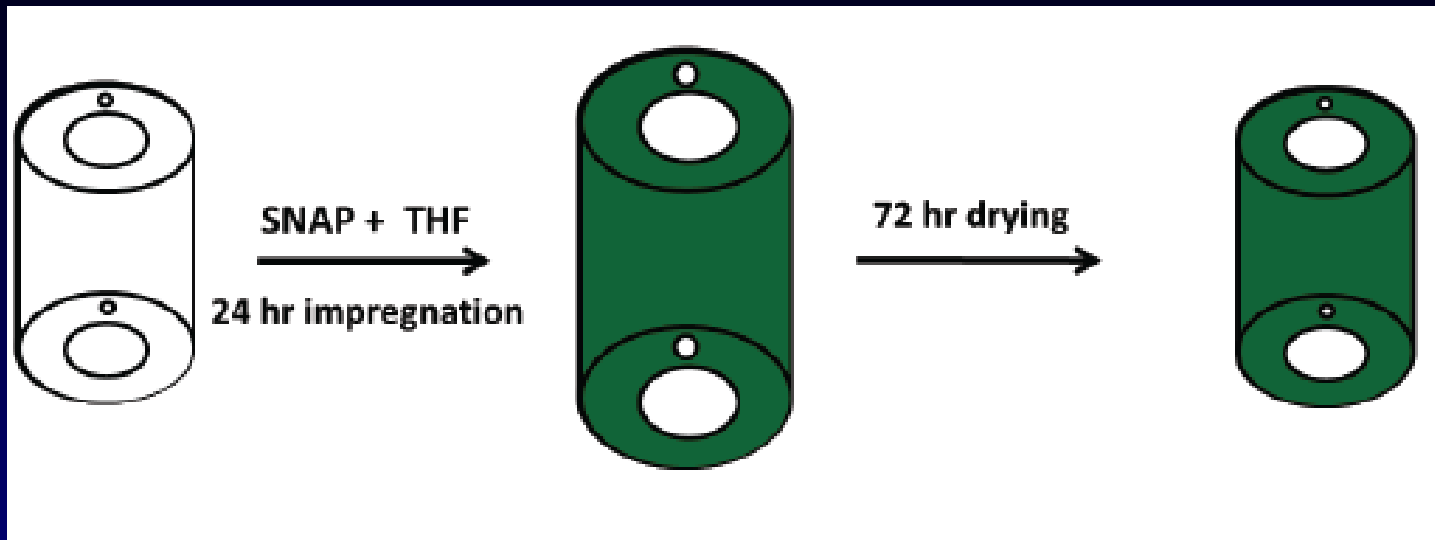
[§]Biocrede Inc., Plymouth, Michigan 48170, United States

S Supporting Information

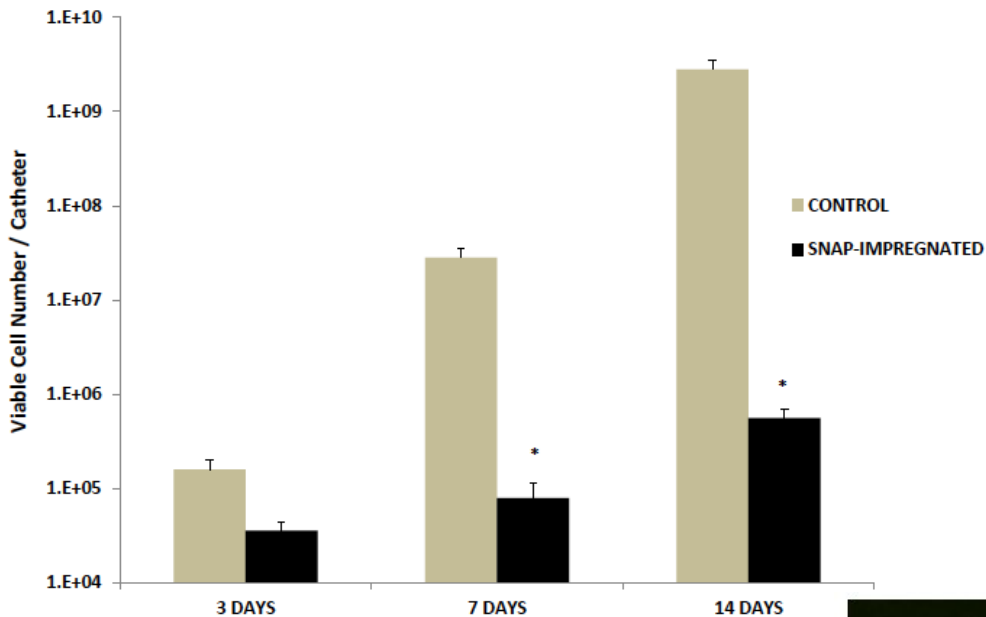
ABSTRACT: Urinary Foley catheters are utilized for management of hospitalized patients and are associated with high rates of urinary tract infections (UTIs). Nitric oxide (NO) potently inhibits microbial biofilm formation, which is the primary cause of catheter associated UTIs (CAUTIs). Herein, commercial silicone Foley catheters are impregnated via a solvent swelling method with S-nitroso-N-acetylpenicillamine (SNAP), a synthetic NO donor that exhibits long-term NO release and stability when incorporated into low water-uptake polymers. The proposed catheters generate NO surface-fluxes $>0.7 \times 10^{-10} \text{ mol min}^{-1} \text{ cm}^{-2}$ for over one month under physiological conditions, with minimal SNAP leaching. These biomedical devices are demonstrated to significantly decrease formation of biofilm on the surface of the catheter tubings over 3, 7, and 14 day periods by microbial species (*Staphylococcus epidermidis* and *Proteus mirabilis*) commonly causing CAUTIs. Toxicity assessment demonstrates that the SNAP-impregnated catheters are fully biocompatible, as extracts of the catheter tubings score 0 on a 3-point grading scale using an accepted mouse fibroblast cell-line toxicity model.



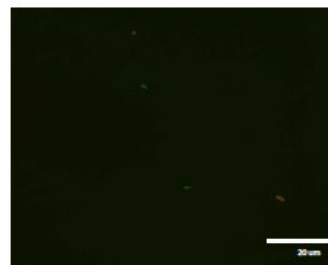
SNAP Impregnation into Catheter Tubing (e.g. Foley Catheters)



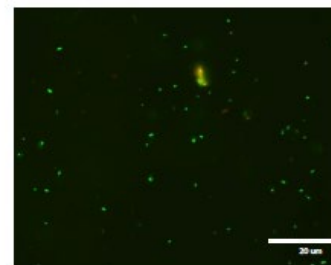
Staphylococcus Epidermidis Biofilm Cell Count



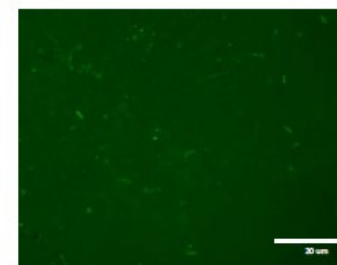
Antimicrobial Activity of SNAP Impregnated Foley Catheters Toward *Staph. epidermis* in CDC Bioreactor for 3, 7, and 14 d



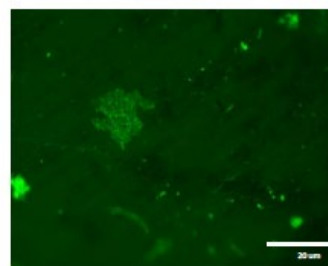
3 Days Biofilm – SNAP Catheter



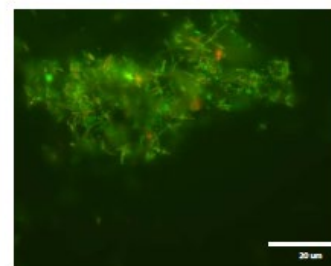
7 Days Biofilm – SNAP Catheter



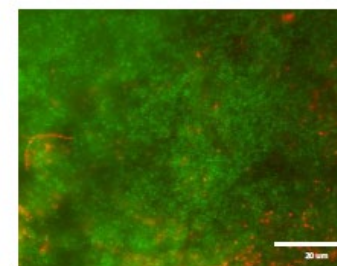
14 Days Biofilm – SNAP Catheter



3 Days Biofilm – Control Catheter

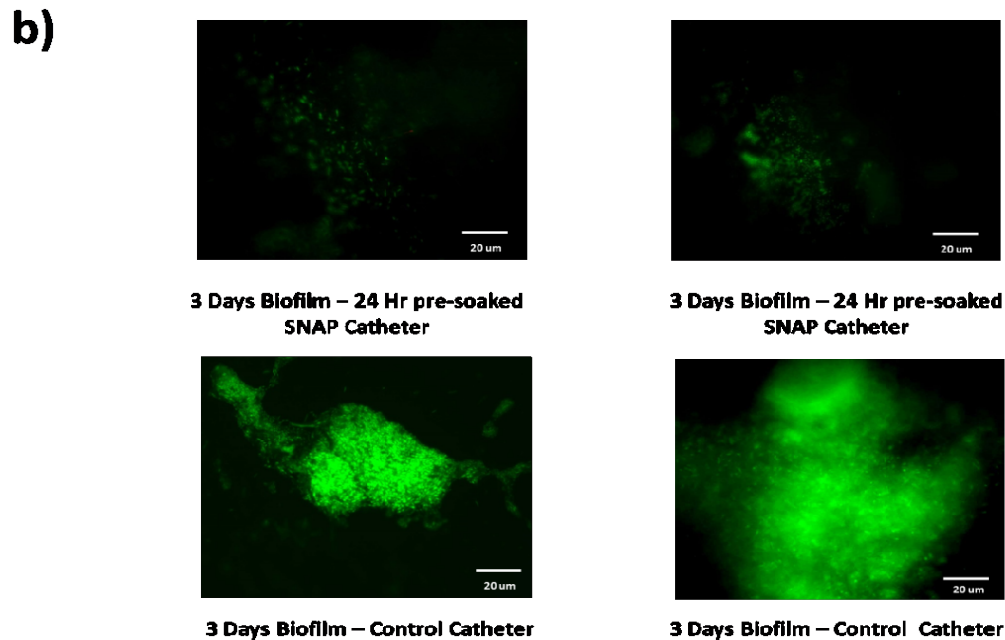
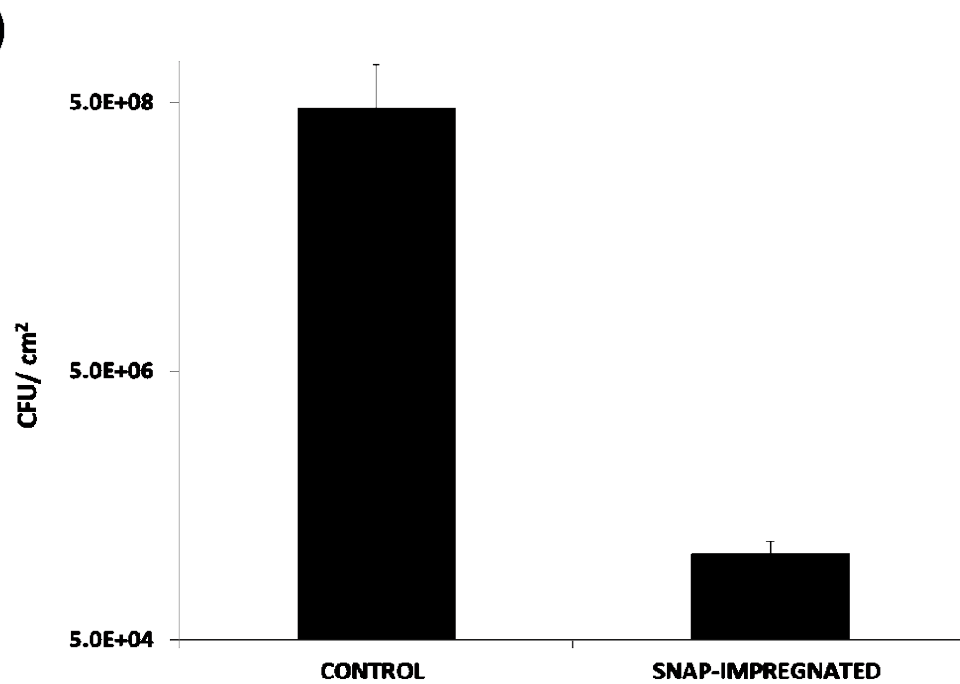


7 Days Biofilm – Control Catheter



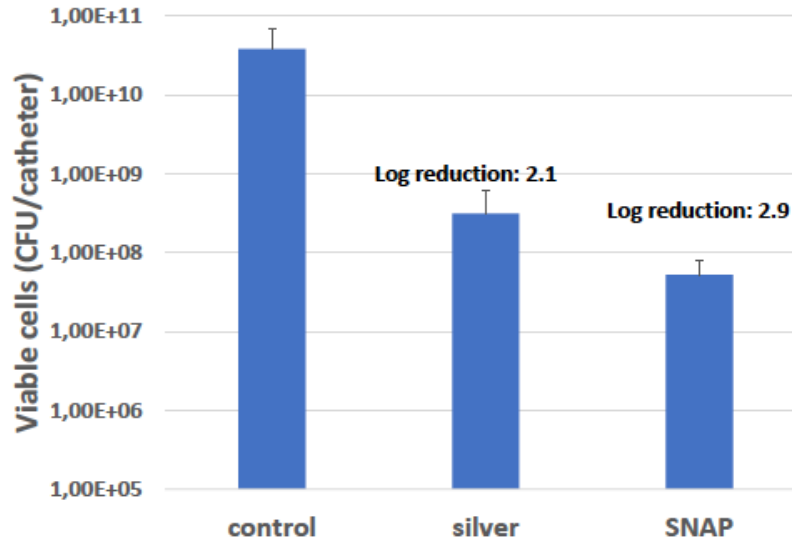
14 Days Biofilm – Control Catheter

Antimicrobial Activity of SNAP Impregnated Commercial Silicone Rubber Foley Catheters Toward *P. mirabilis* in CDC Bioreactor for 3 d

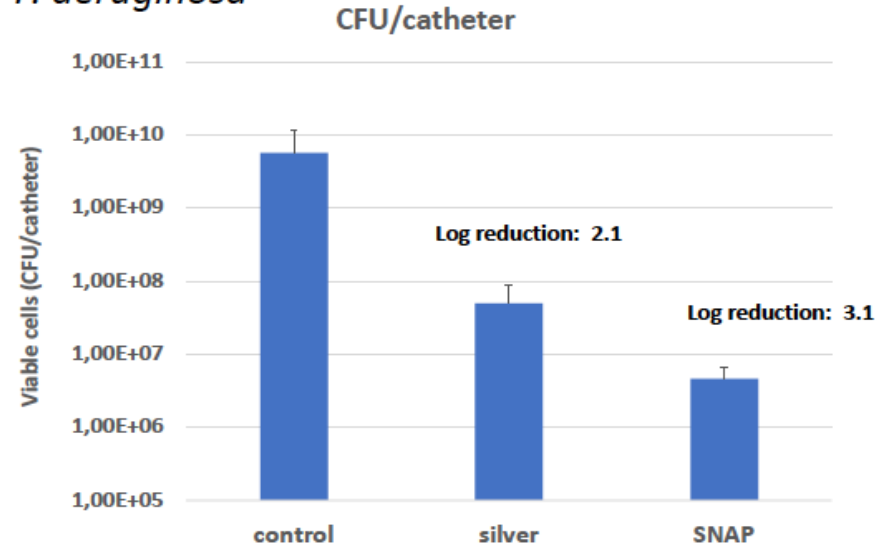


Comparison of Commercial Ag⁰ Particle Impregnated Urinary Catheters vs. New SNAP-Doped Catheters

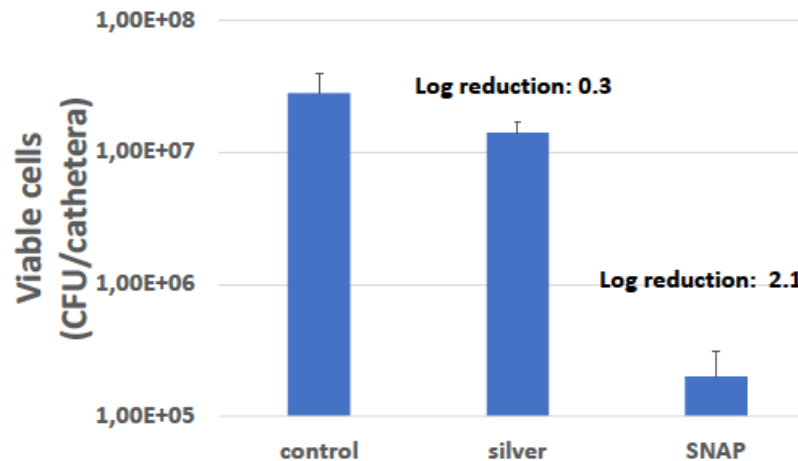
Proteus mirabilis



P. aeruginosa



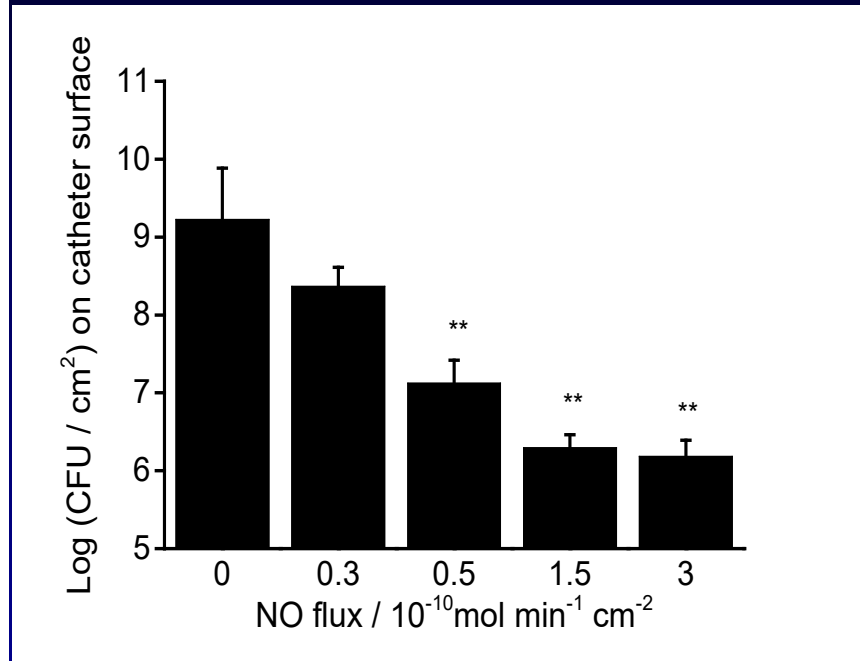
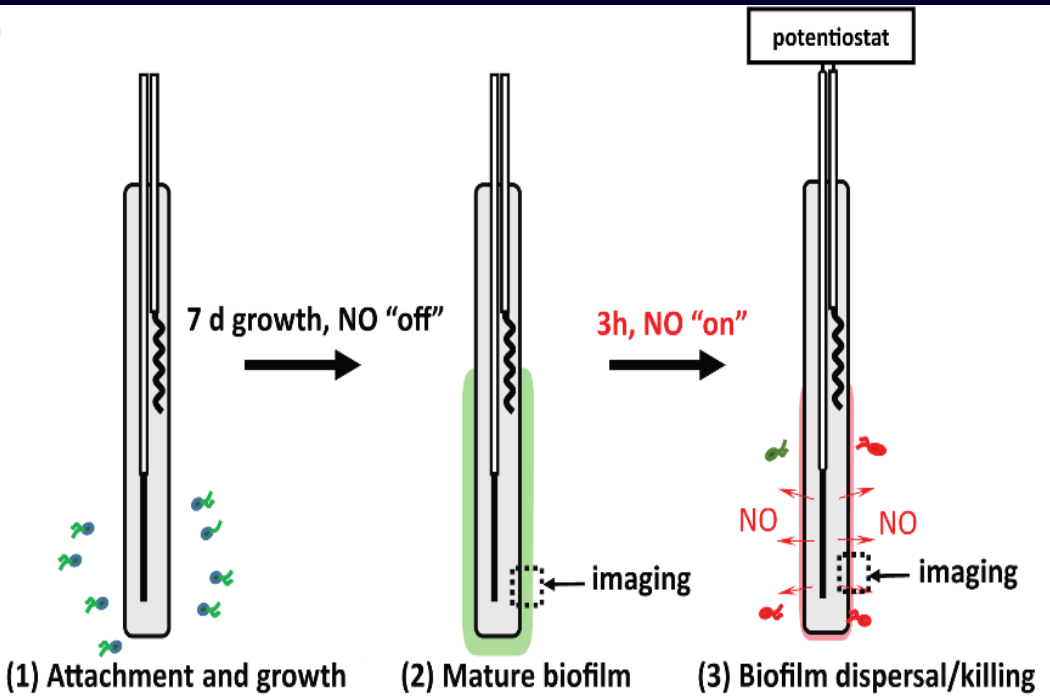
Escherichia coli



7-day
experiments
within CDC
bioreactor

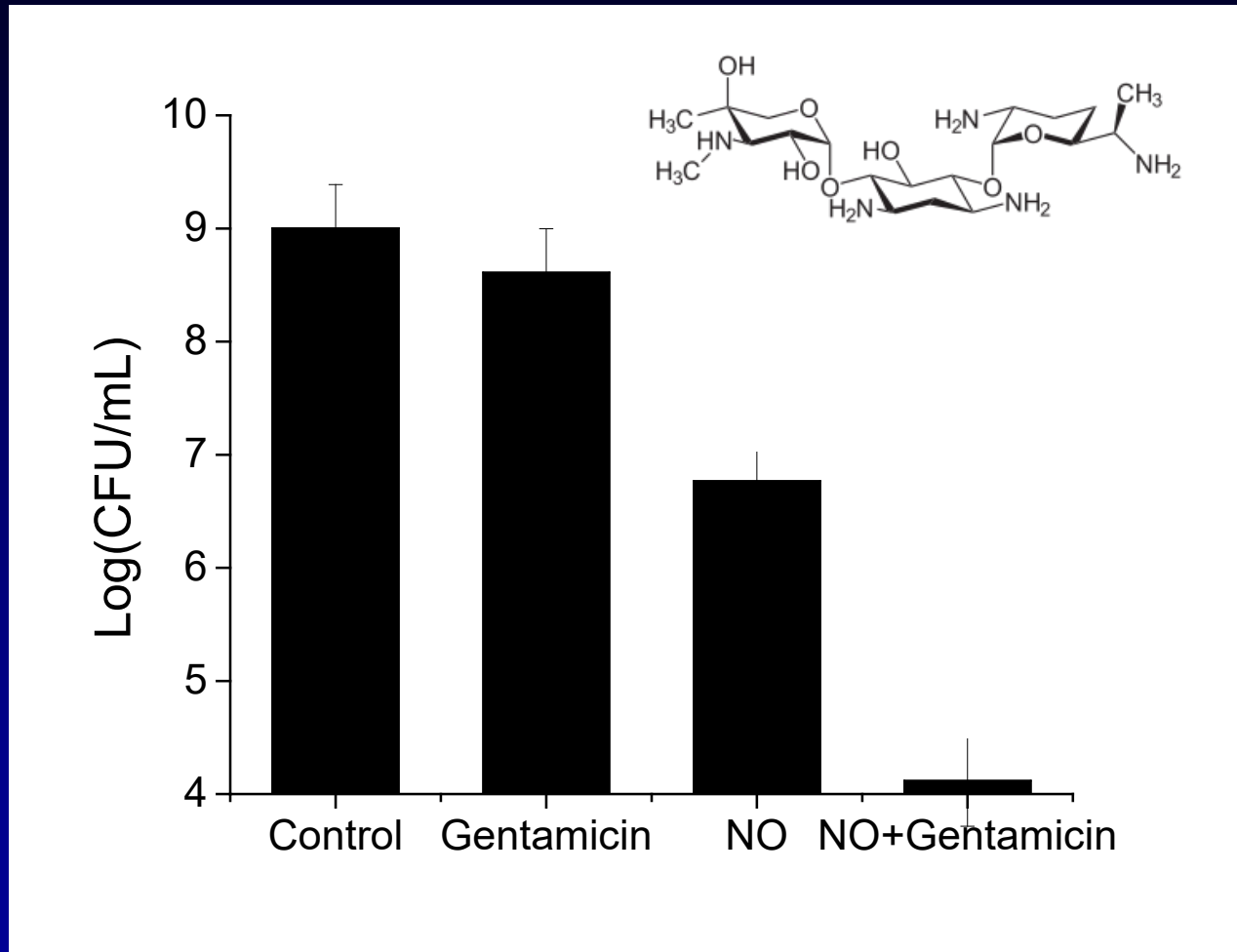
Note:
NO release
Catheter only
had 4 wt% SNAP

Removal of Mature Biofilms



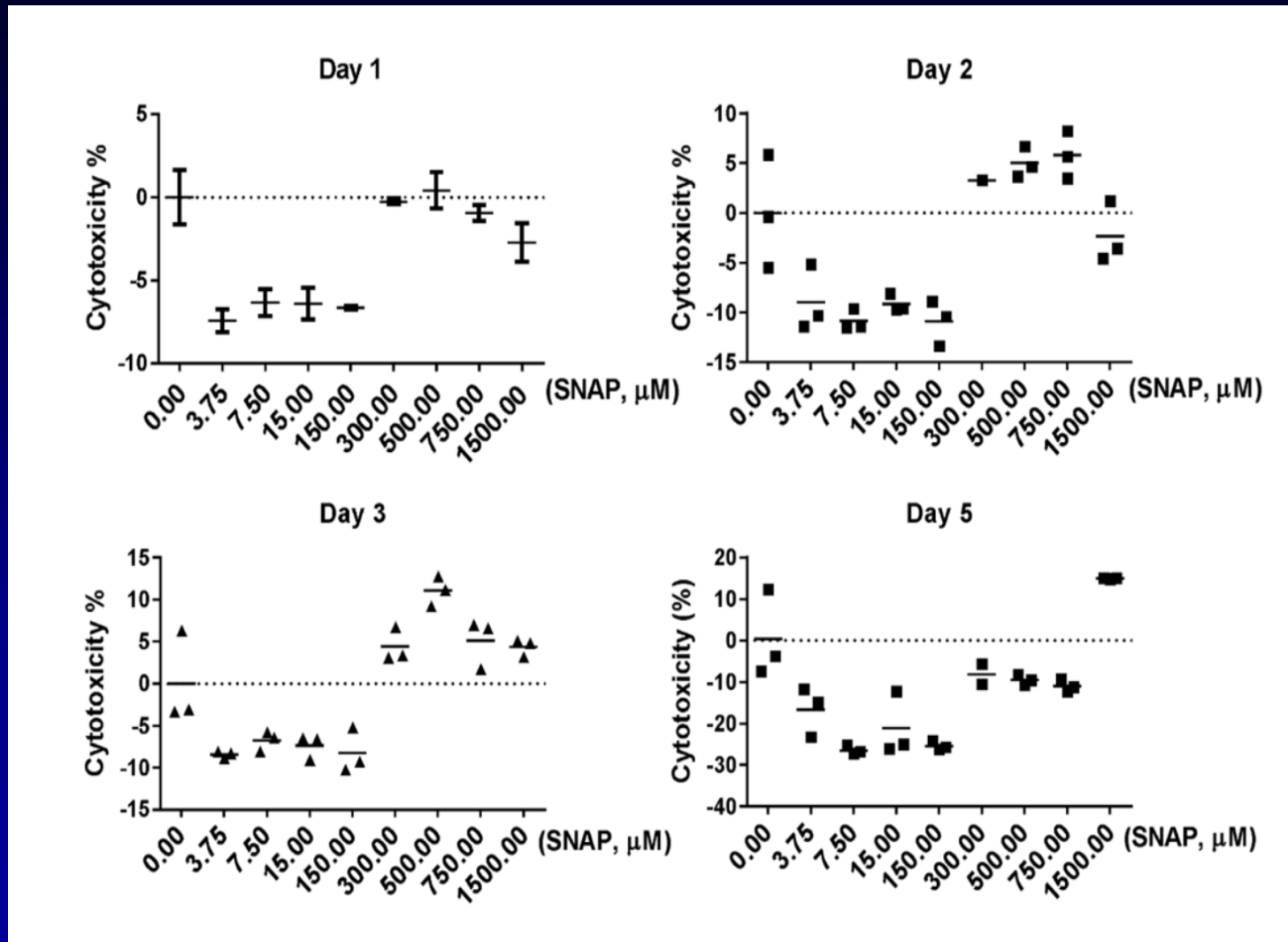
P. aeruginosa biofilm grown for 7 d before any treatment

Synergy of NO with Antibiotic Treatment



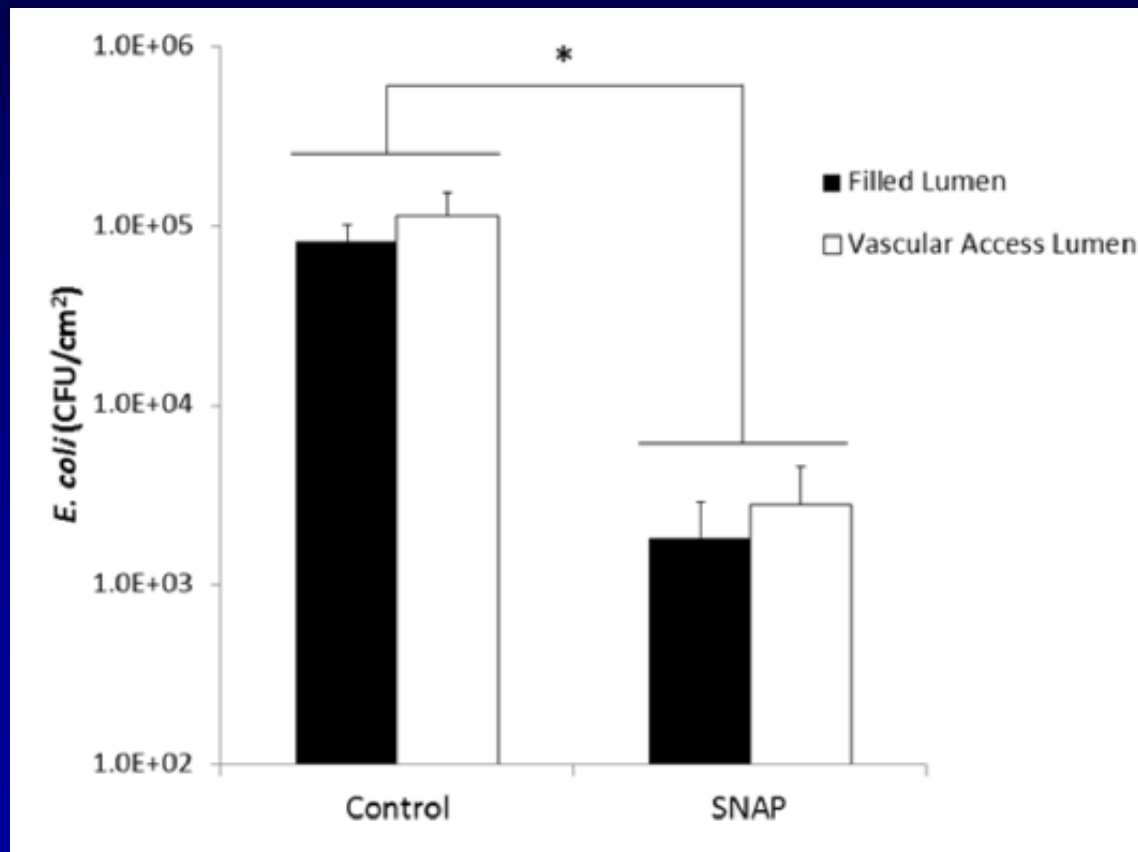
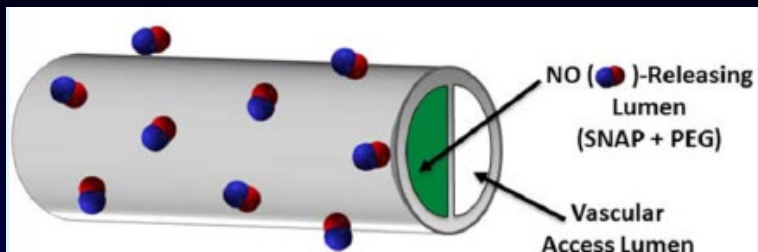
NO release + antibiotic (100 $\mu\text{g}/\text{mL}$) treatment for 3 h on 7 d *P. aeruginosa* biofilms

Cytotoxicity Study of SNAP on Human Cells (AD-293 cells—Embryonic Kidney Cell line)

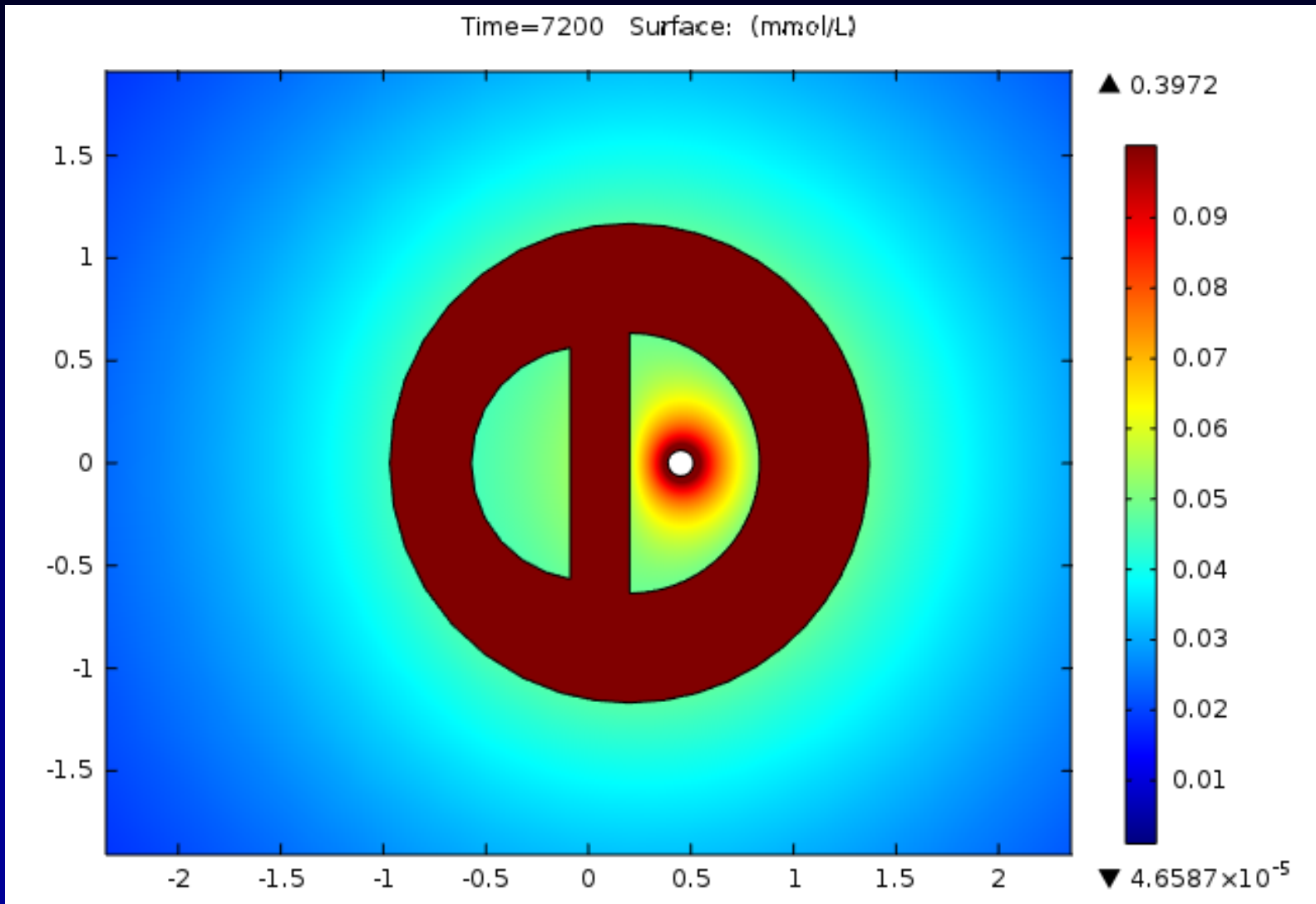


Toxicity assay based on LDH activity measurements

Alternate Configuration: Filling and Sealing One Lumen of Dual Lumen Catheters – Nothing Comes Out Except NO



Finite Element Simulations of NO Concentration Profile in Dual Lumen Catheter Configuration (Comsol Software)



$$\text{Partition coefficient } K = S_{\text{silicone}} / S_{\text{water}} = 7$$
$$D_{\text{NO}}(\text{polymer}) = D_{\text{NO}}(\text{water})$$

NO-Release Collaborators/Support

Current/Recent Graduate Students/Undergraduates/Post-Docs

Alex Wolf	Dipankar Koley	Wenyi Cai
Hang Ren	Elizabeth Brisbois	Yaqi Wo
Zheng Zheng	Hitesh Handa	Wen Wen
Gergely Lautner	Xuwei Wang	Kyoung-Ha Cha
Alessandro Colletta	Woonghee Lee	Alex Ketchum
Andrew Hunt	Jianfeng Wu	Joanna Zajda
Bo Peng	Melissa Reynolds	Josh Doverspike
Kamila Konopinska	Qi Zhang	
Yu Qin	Orsi Lautner	

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- NIH: EB-023294
- NIH: HD-087071
- NIH: EB-024038
- JDRF / NIH: EB-019566
- MC3 Inc./ NOTA Labs
- BICI/ Beijing-Epione
- Instrumentation Laboratory

Faculty/Other

Nicolai Lehnert (U of M)
Prabir Roy-Chaudhury (U of Cinc.)
Robert Bartlett (UM-Surgery)
Chuanwu Xi (UM-SPH)
Yu Qin (U of M)