

Design of Catheter Coatings for Localized Release of Antimicrobial Peptide Mimetics

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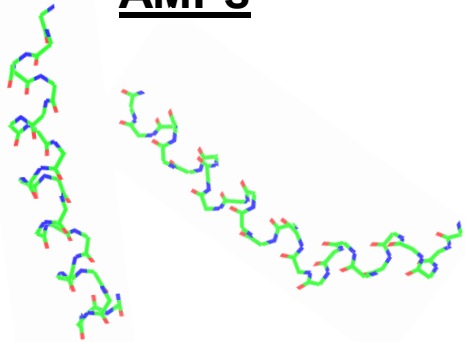


β -peptide mimetics of antimicrobial peptides

Antimicrobial α -peptides

- Components of the innate immune system
- Kill bacterial and fungal pathogens
- Membrane disruption based mechanisms: more difficult for pathogens to develop resistance

AMPs



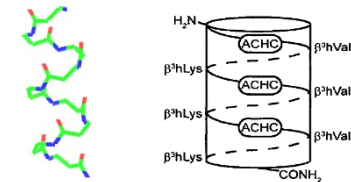
Limitations

- Degradation by cellular proteases
- Low activity at physiologic ionic strength

Antimicrobial β -peptides

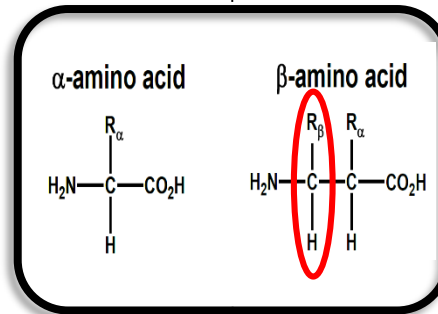
- Oligomers of β -amino acids
- Peptidomimetics of AMPs
- Can be designed to fold into specific secondary structures

β -Peptide

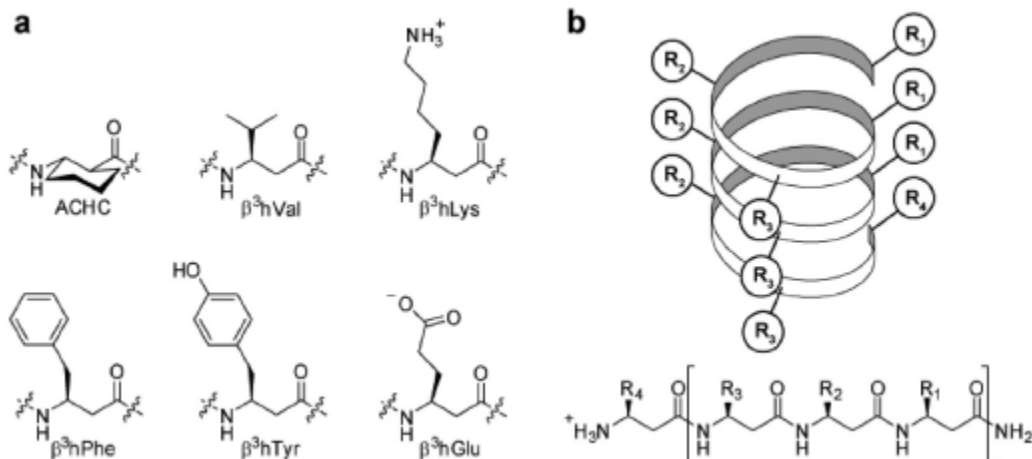


Advantages

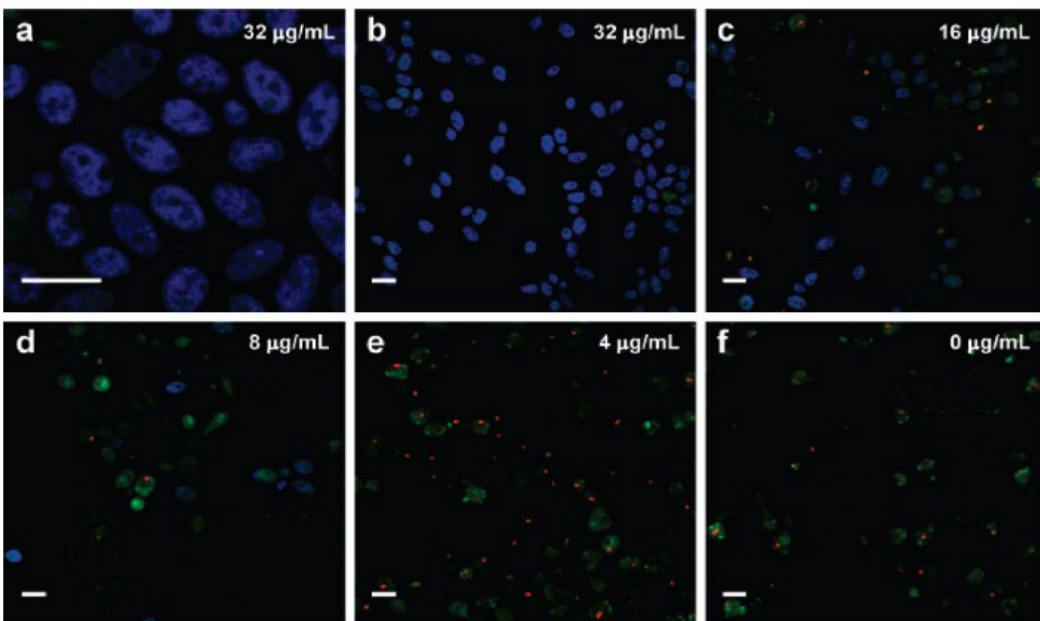
- Stable in physiologic conditions, high ionic strength and against proteolytic enzymes



Antifungal β -peptides

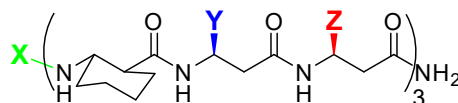
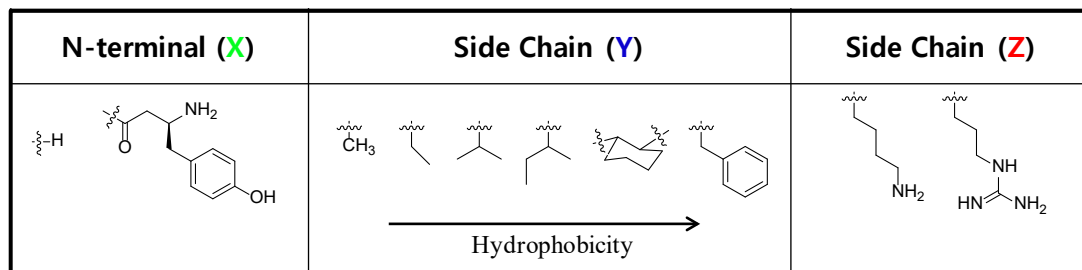
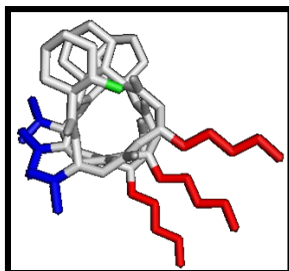


- Antifungal β -peptide
 - Globally amphiphillic
 - Length \rightarrow 9-mer, 10-mer
 - Charge \rightarrow +3, +4
- Identified β -peptides with antifungal activity against *C. albicans* and selective (non-toxic towards hRBCs)
- Fluorescently labeled peptide for tracking

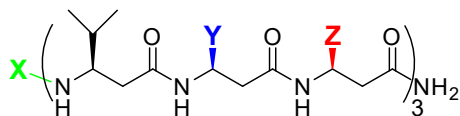


Blue – Labeled peptide
Green – Cytoplasm stain
Red – Intra-vacuolar structures
(only in metabolic active cells)

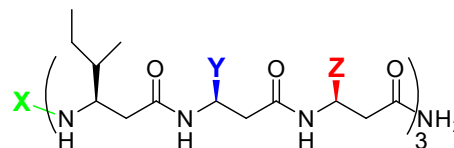
Effect of hydrophobicity and helicity on β -peptide activity and selectivity



- | | |
|---|--|
| 1 : X = H, Y = β^3 -hAla, Z = β^3 -hLys | 9 : X = β^3 -hTyr, Y = β^3 -hAla, Z = β^3 -hLys |
| 2 : X = H, Y = β^3 -Et, Z = β^3 -hLys | 10 : X = β^3 -hTyr, Y = β^3 -Et, Z = β^3 -hLys |
| 3 : X = H, Y = β^3 -Et, Z = β^3 -hArg | 11 : X = β^3 -hTyr, Y = β^3 -Et, Z = β^3 -hArg |
| 4 : X = H, Y = β^3 -hVal, Z = β^3 -hLys | 12 : X = β^3 -hTyr, Y = β^3 -hVal, Z = β^3 -hLys |
| 5 : X = H, Y = β^3 -hVal, Z = β^3 -hArg | 13 : X = β^3 -hTyr, Y = β^3 -hVal, Z = β^3 -hArg |
| 6 : X = H, Y = ACHC, Z = β^3 -hLys | 14 : X = β^3 -hTyr, Y = ACHC, Z = β^3 -hLys |
| 7 : X = H, Y = ACHC, Z = β^3 -hArg | 15 : X = β^3 -hTyr, Y = ACHC, Z = β^3 -hArg |
| 8 : X = H, Y = β^3 -hPhe, Z = β^3 -hLys | 16 : X = β^3 -hTyr, Y = β^3 -hPhe, Z = β^3 -hLys |

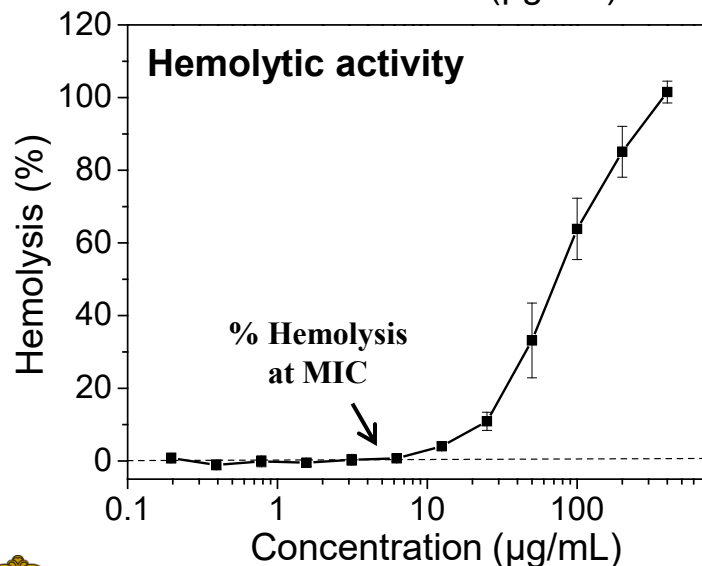
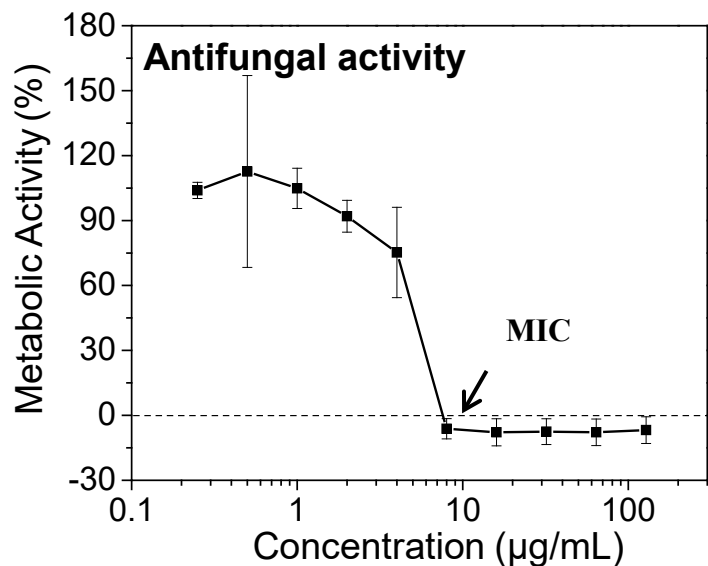


- | |
|--|
| 17 : X = H, Y = β^3 -hVal, Z = β^3 -hLys |
| 18 : X = H, Y = β^3 -hVal, Z = β^3 -hArg |
| 19 : X = β^3 -hTyr, Y = β^3 -Et, Z = β^3 -hArg |
| 20 : X = β^3 -hTyr, Y = β^3 -hVal, Z = β^3 -hLys |
| 21 : X = β^3 -hTyr, Y = β^3 -hVal, Z = β^3 -hArg |



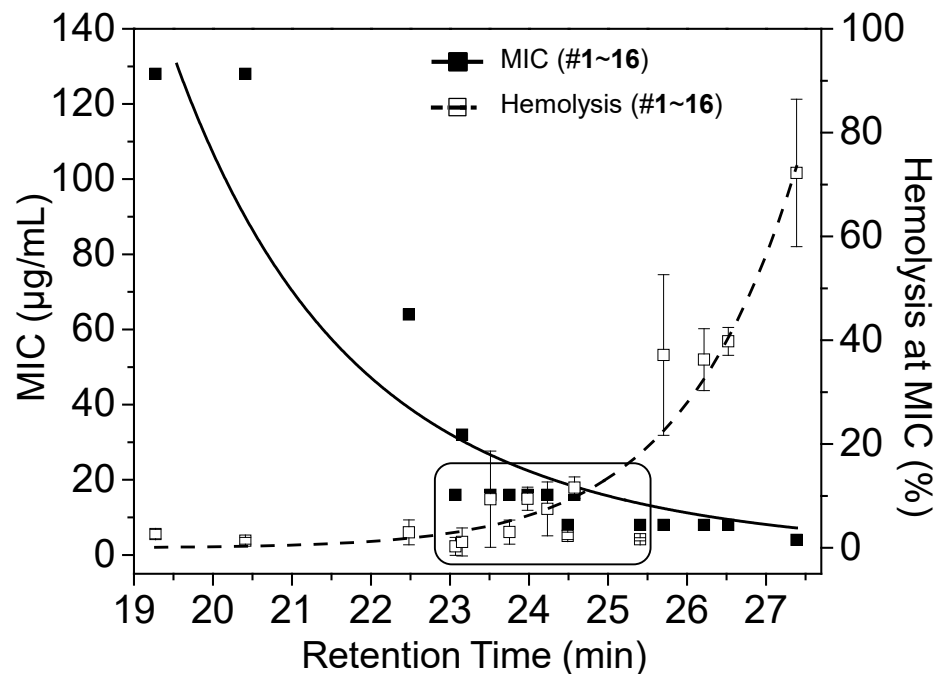
- | |
|--|
| 22 : X = H, Y = β^3 -Et, Z = β^3 -hLys |
| 23 : X = H, Y = β^3 -Et, Z = β^3 -hArg |
| 24 : X = β^3 -hTyr, Y = β^3 -Et, Z = β^3 -hLys |
| 25 : X = β^3 -hTyr, Y = β^3 -Et, Z = β^3 -hArg |

β -peptide antifungal and hemolytic activity



β -Peptide	RT (min \pm SD)	MIC (μ g/mL)	% Hemolysis at MIC \pm SD
1	19.3 \pm 0.1	> 128	2.6 \pm 0.9*
2	22.5 \pm 0.2	64	3.0 \pm 2.4
3	23.2 \pm 0.1	32	1.1 \pm 2.7
4	24.5 \pm 0.2	8	2.3 \pm 0.7
5	25.4 \pm 0.1	8	1.6 \pm 0.3
6	23.1 \pm 0.2	16	0.3 \pm 1.7
7	23.8 \pm 0.1	16	3.0 \pm 2.3
8	26.2 \pm 0.2	8	36.4 \pm 6.0
9	20.4 \pm 0.2	128	1.4 \pm 0.6
10	23.5 \pm 0.1	16	9.4 \pm 9.3
11	24.2 \pm 0.1	16	7.5 \pm 5.2
12	25.7 \pm 0.1	8	37 \pm 15
13	26.5 \pm 0.2	8	39.8 \pm 2.7
14	24.0 \pm 0.2	16	9.5 \pm 2.2
15	24.6 \pm 0.2	16	11.6 \pm 2.1
16	27.4 \pm 0.2	4	72 \pm 14
17	22.5 \pm 0.1	128	2.8 \pm 0.1
18	23.5 \pm 0.1	64	0.9 \pm 1.9
19	22.7 \pm 0.2	128	3.2 \pm 2.9
20	24.3 \pm 0.2	32	8.8 \pm 3.6
21	25.2 \pm 0.2	16	4.2 \pm 2.0
22	22.8 \pm 0.2	> 128	3.1 \pm 4.4*
23	23.8 \pm 0.1	128	4.5 \pm 3.2
24	24.6 \pm 0.1	32	7.2 \pm 5.0
25	25.7 \pm 0.2	16	7.2 \pm 3.4

β -peptide hydrophobicity correlates with antifungal and hemolytic activity

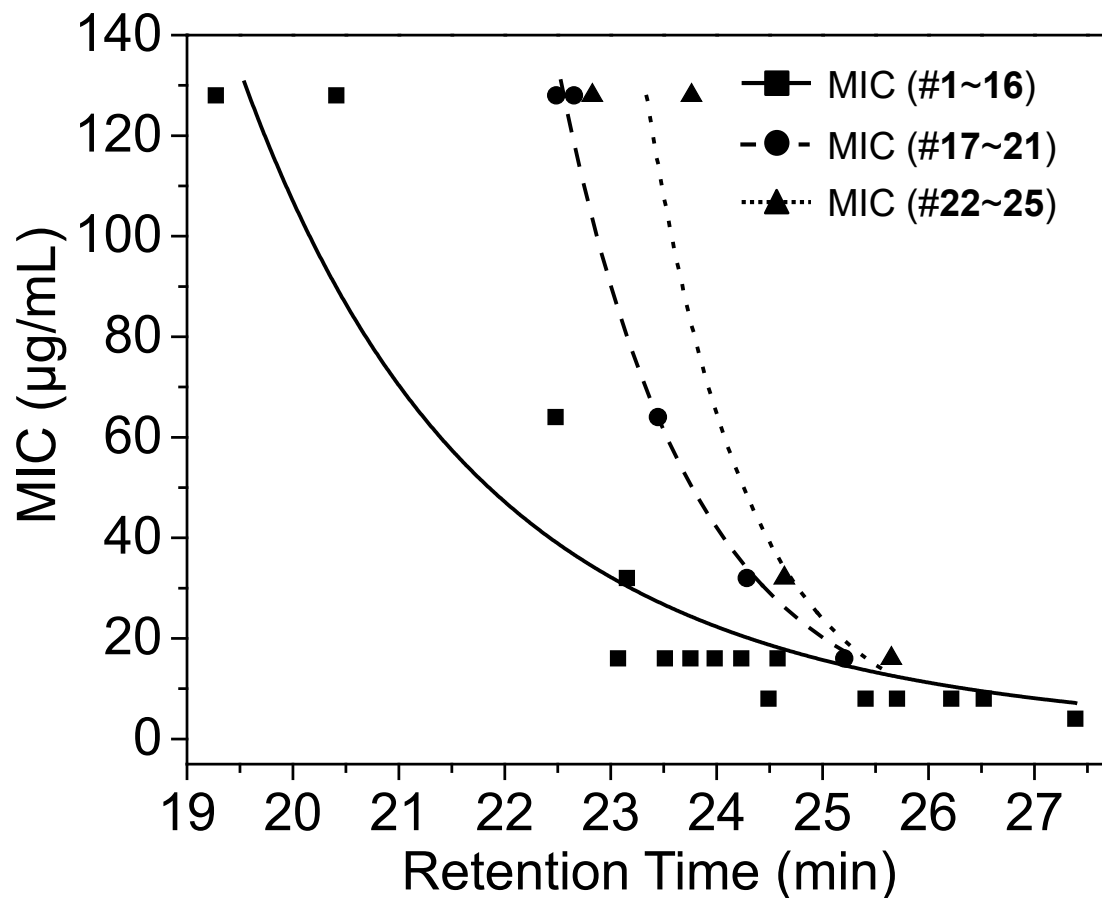


Peptide #	RT ^a (min)	MIC ^b ($\mu\text{g/mL}$)		
		ATCC90028	K1	SC5314
1	19.3	>128	>128	>128
2	22.5	64	64	64
6	23.1	32	32	16
3	23.2	32	32	32
7	23.8	32	16	16
4	24.5	16	16	8
5	25.4	16	8	8
8	26.2	8	4	8

➤ Window of hydrophobicity where β -peptides have selective antifungal activity

➤ β -Peptide planktonic MICs were similar across multiple *C. albicans* strains

β -peptide helicity affects antifungal activity

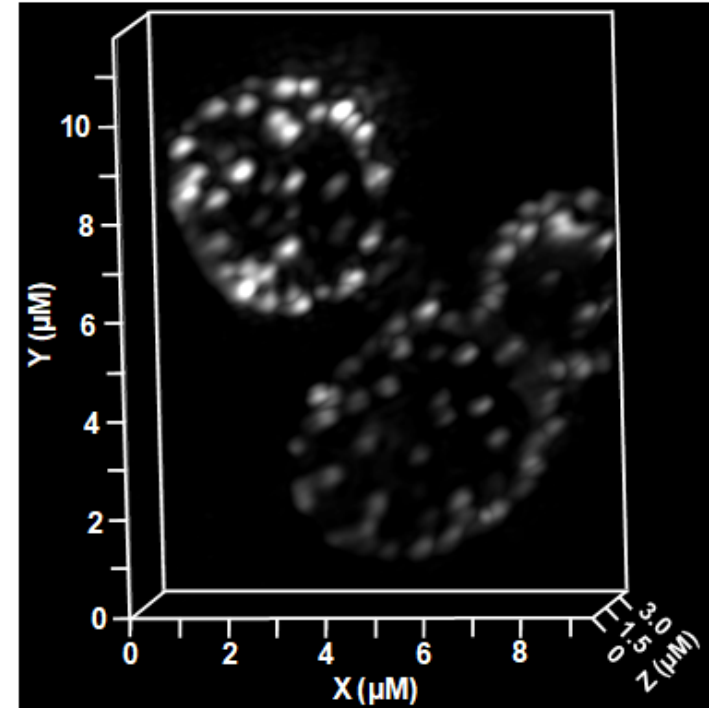
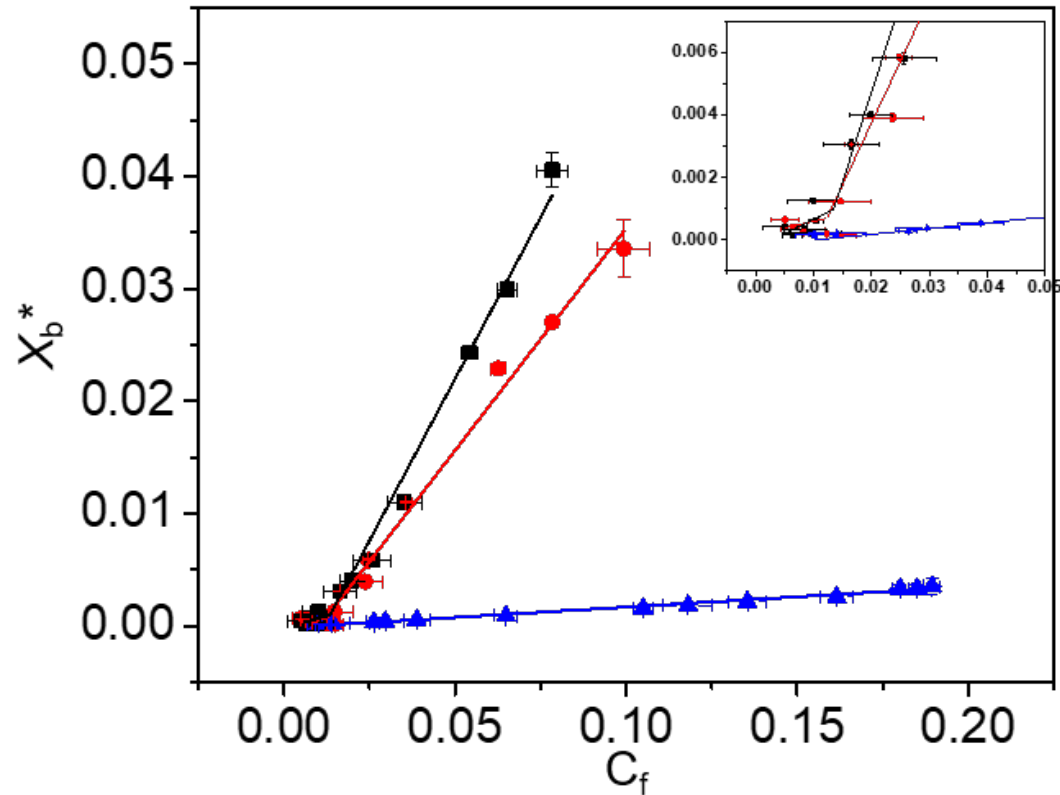


1-16: 3 ACHC side chains
17-25: no ACHC

➤ ACHC incorporation increases antifungal activity in more hydrophobic β -peptides

β -peptides form pore-like structures in the plasma membrane

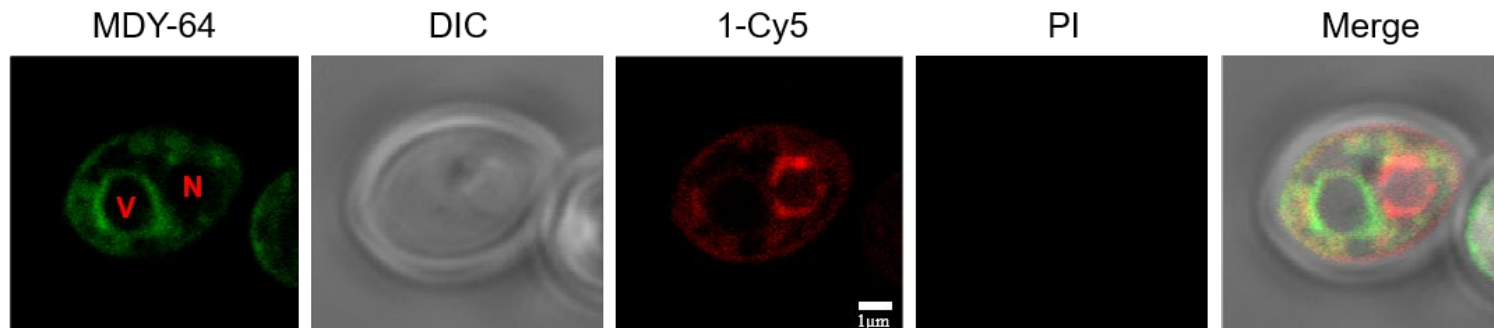
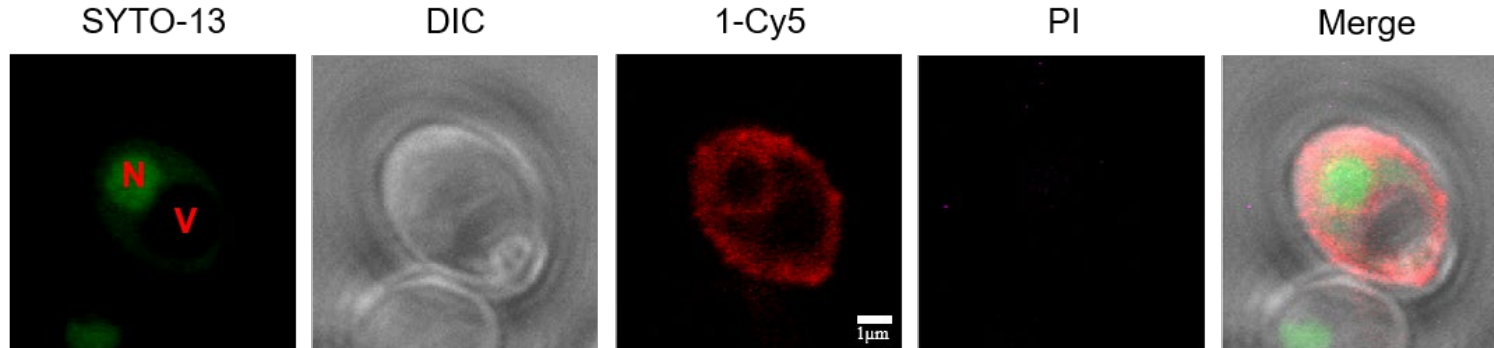
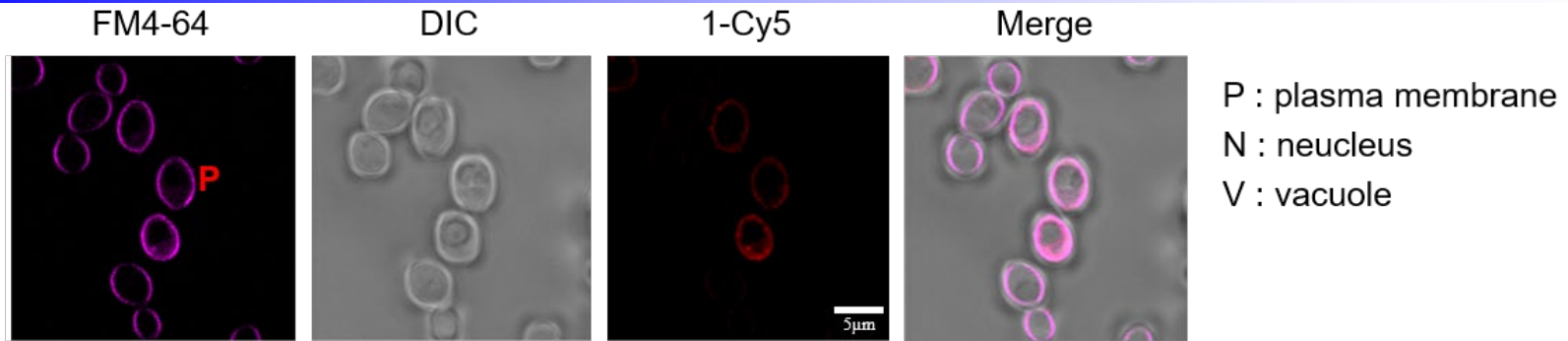
PC : PE (60 : 40)



Cooperative binding of β -peptides to synthetic vesicles

β -peptides form 300 nm aggregates on the surface of *C. albicans* cells

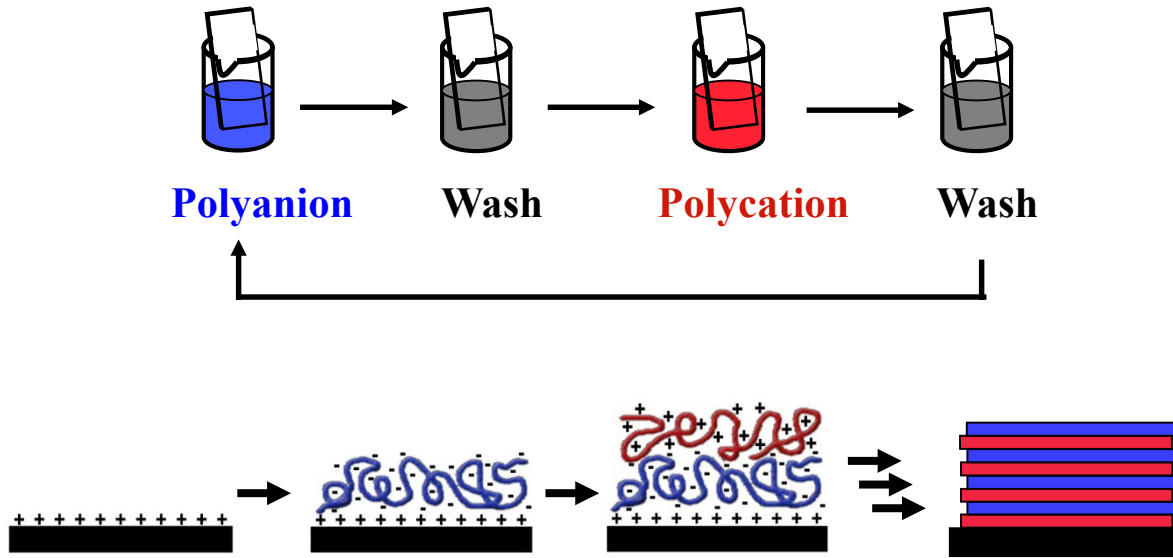
β -peptides disrupt intracellular organelles



β -peptides disrupt the cell membrane, enter the cell and lyse the nucleus and vacuole

Polyelectrolyte multilayer films

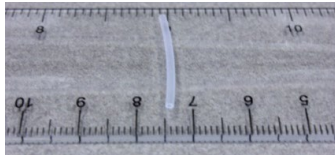
- Layer-by-layer adsorption of oppositely charged polyelectrolytes



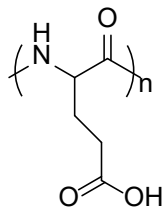
- Electrostatic self assembly process
- Compatible with biomolecules
- Versatile, inexpensive
- Coat topologically complex substrates

PEM fabrication in catheters

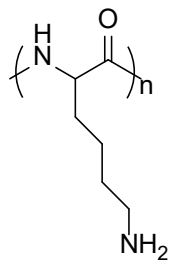
Catheter Segment



860 μm id polyethylene tube



Poly(L-glutamic acid)
(PGA)



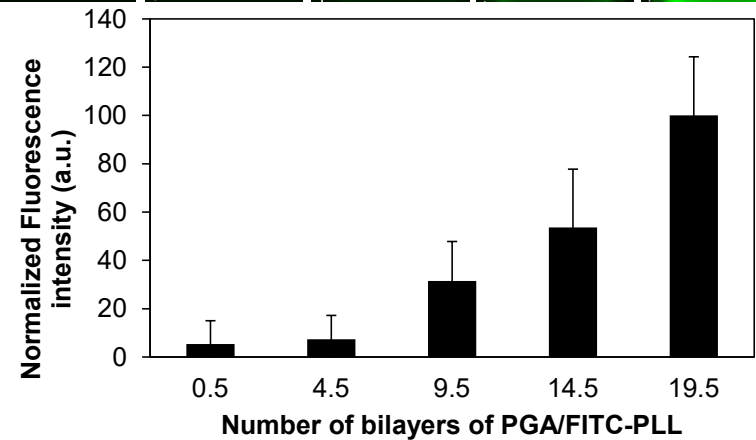
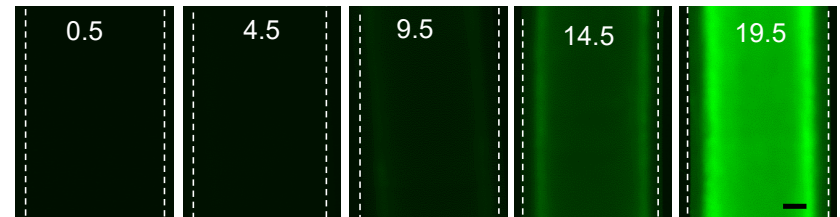
Poly(L-lysine)
(PLL)



Tube

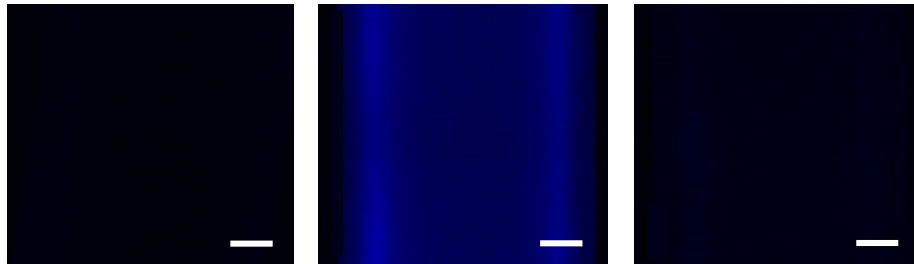
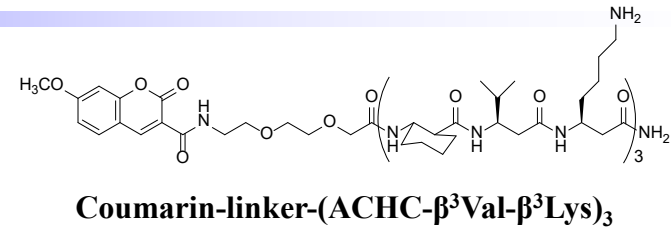
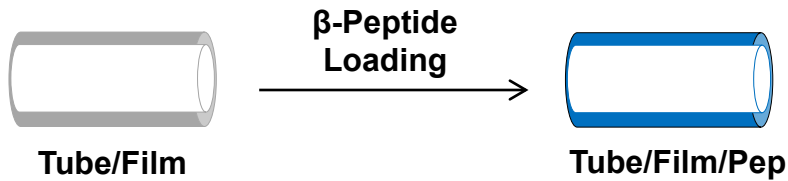
Tube/Film

Bilayer #

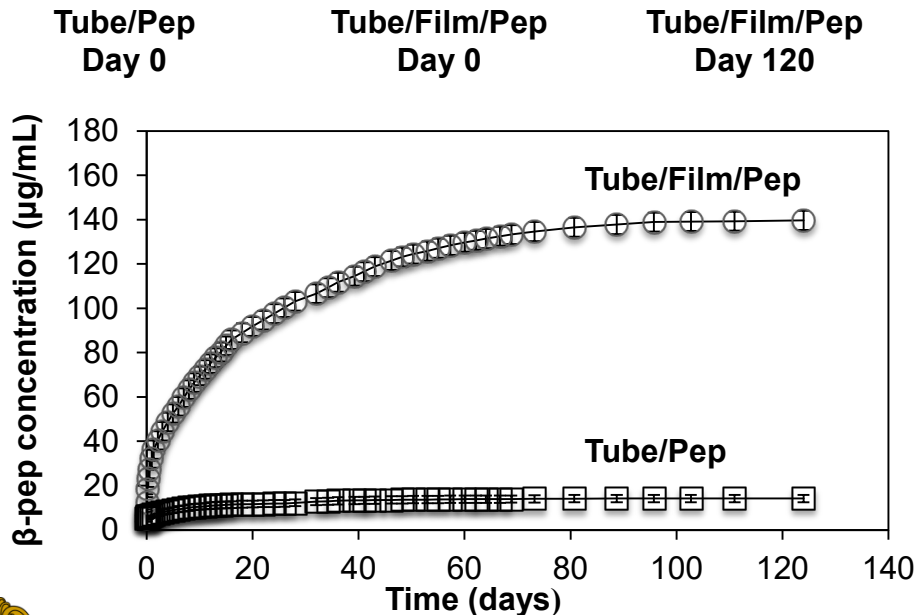


- Supra-linear PEM film growth of PGA/PLL multilayers fabricated on the inner surfaces of catheter tubes

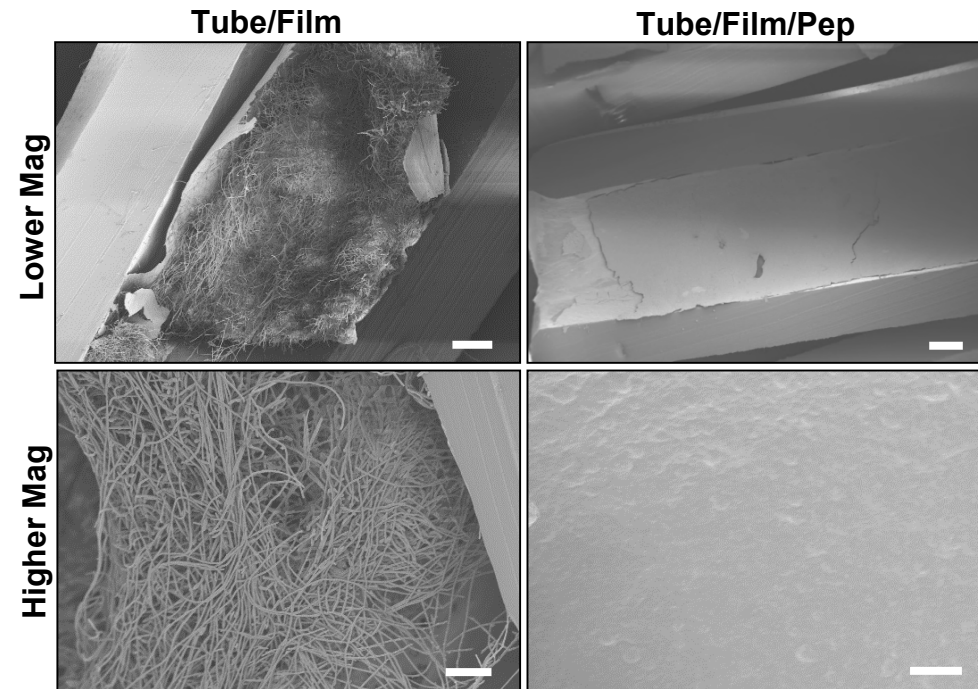
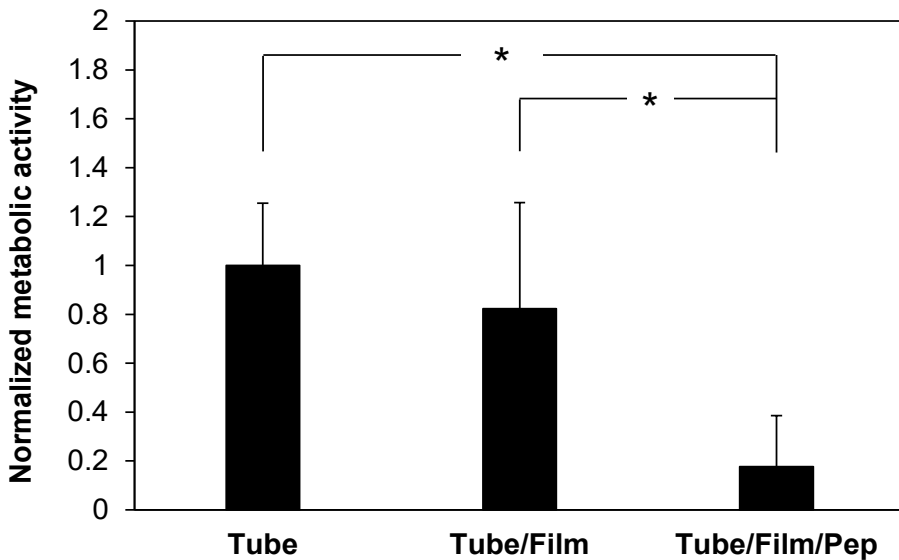
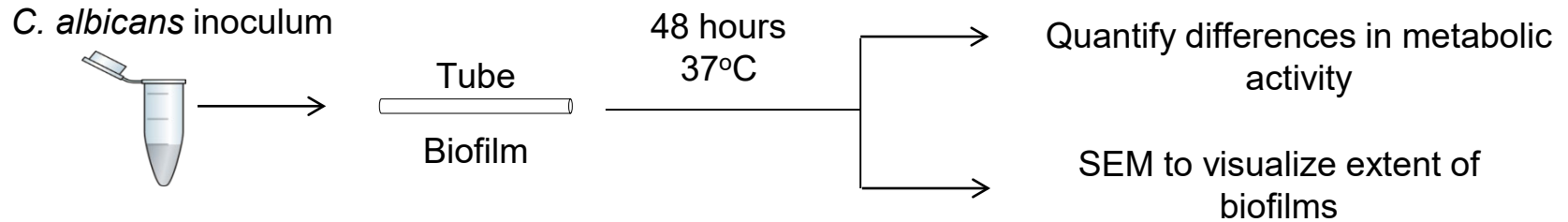
Loading and release of β -peptide from PEM-coated catheters



- β -Peptide can be loaded post PEM fabrication
- Multilayer film enhances the localization of the peptide on the inner surface of the catheter
- β -Peptide is gradually released from film coated, peptide loaded catheters over the course of 60-80 days.



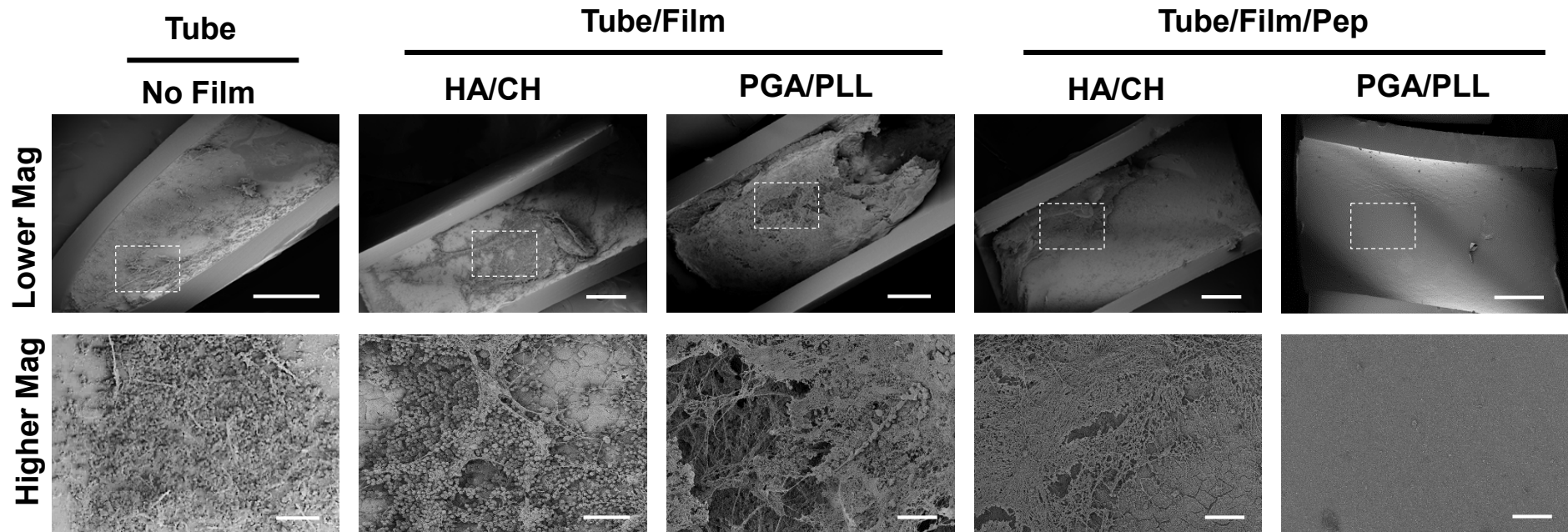
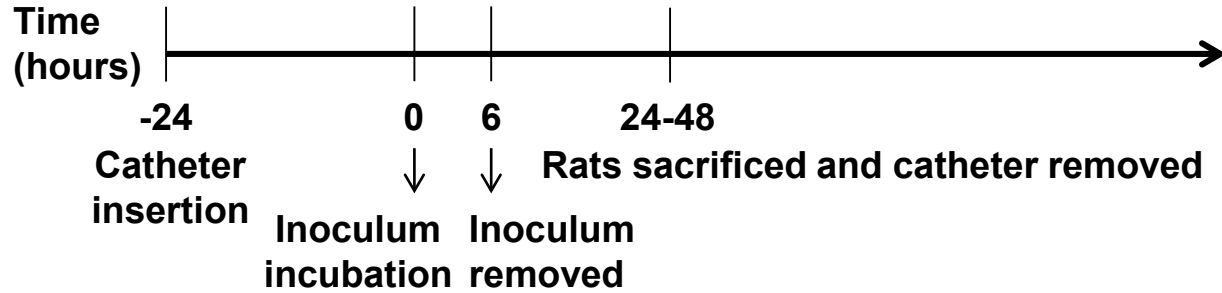
Prevention of *C. albicans* biofilms in β -peptide-loaded catheters



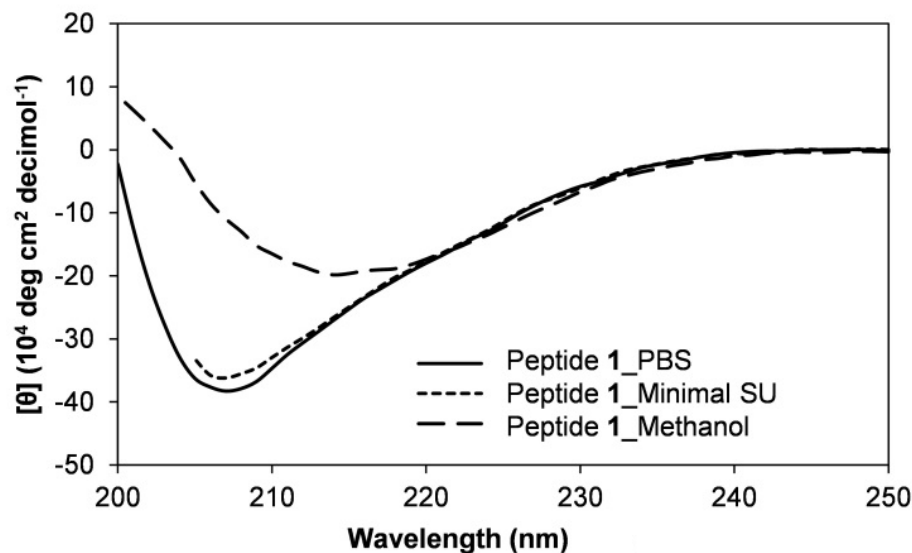
➤ Catheters coated with β -peptide-loaded films inhibit *C. albicans* biofilm formation

Reduction of biofilm formation *in vivo* on β -peptide loaded catheters

- Rat central venous catheter model

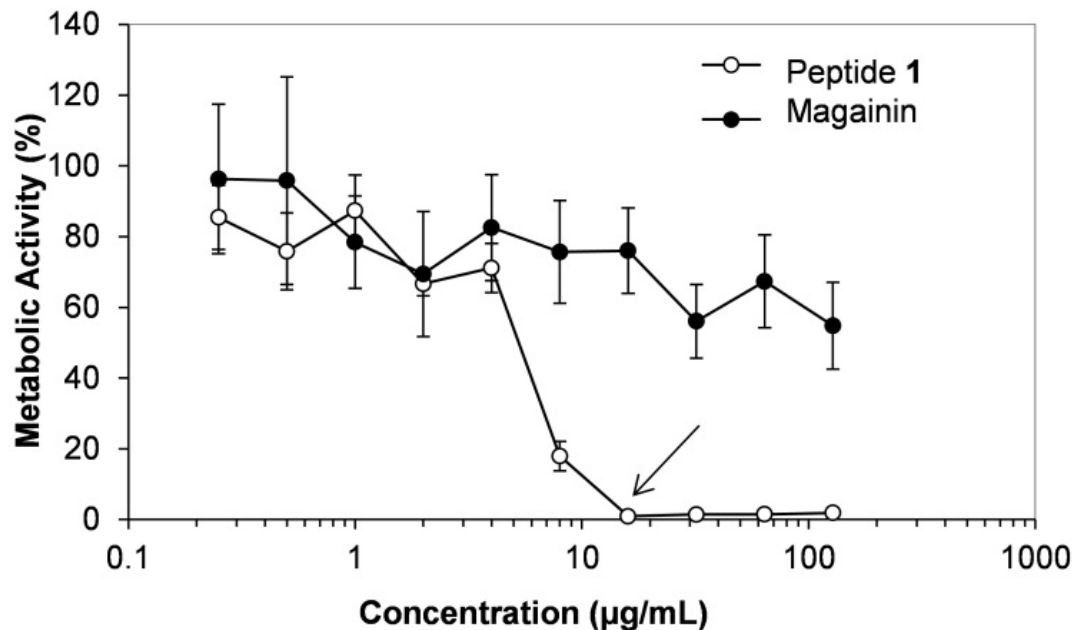


β -peptide activity in synthetic urine



β -peptide remains helical in synthetic urine

MIC=16 $\mu\text{g/ml}$ (8 in RPMI)
MBPC=32 $\mu\text{g/ml}$ (32 in RPMI)

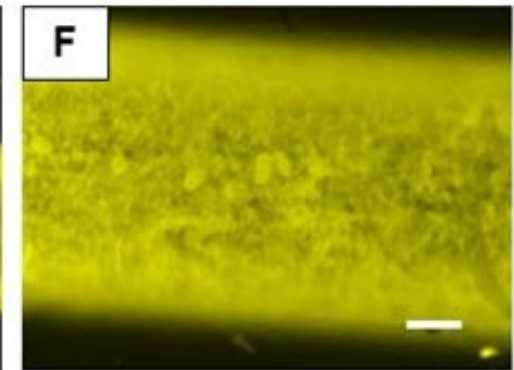
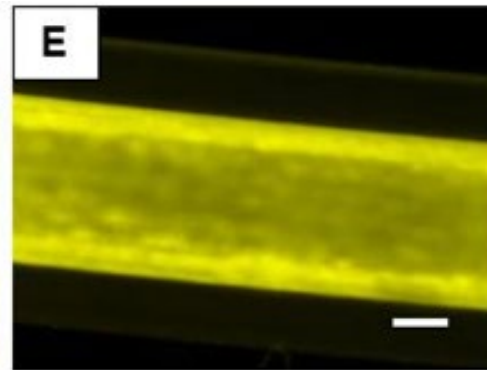
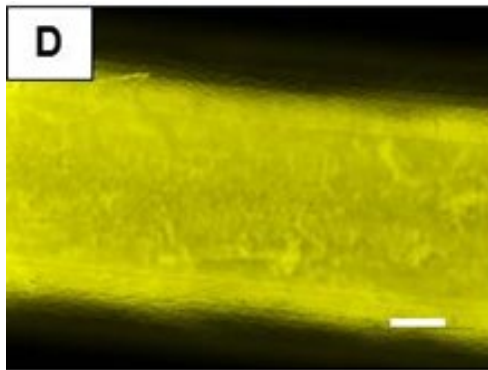


β -peptide release from films on urinary catheters

polyethylene

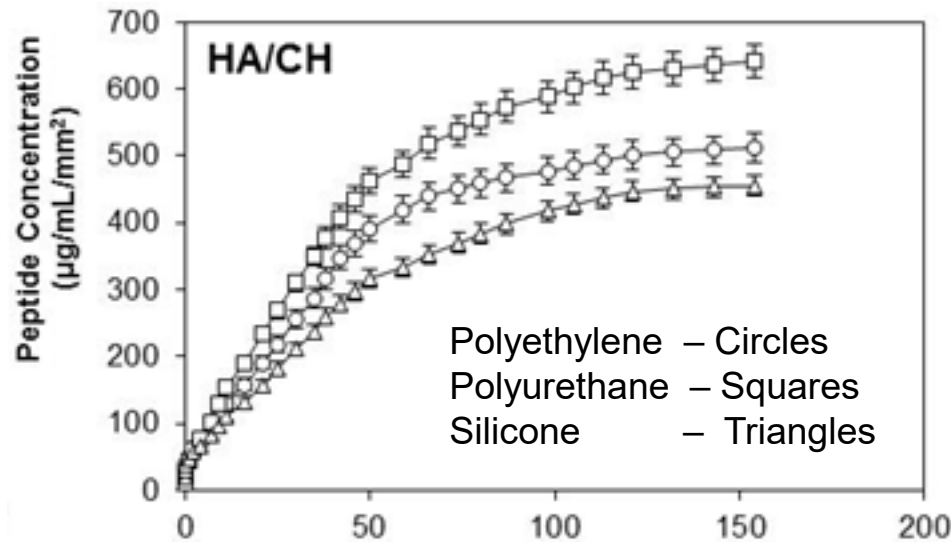
polyurethane

silicone



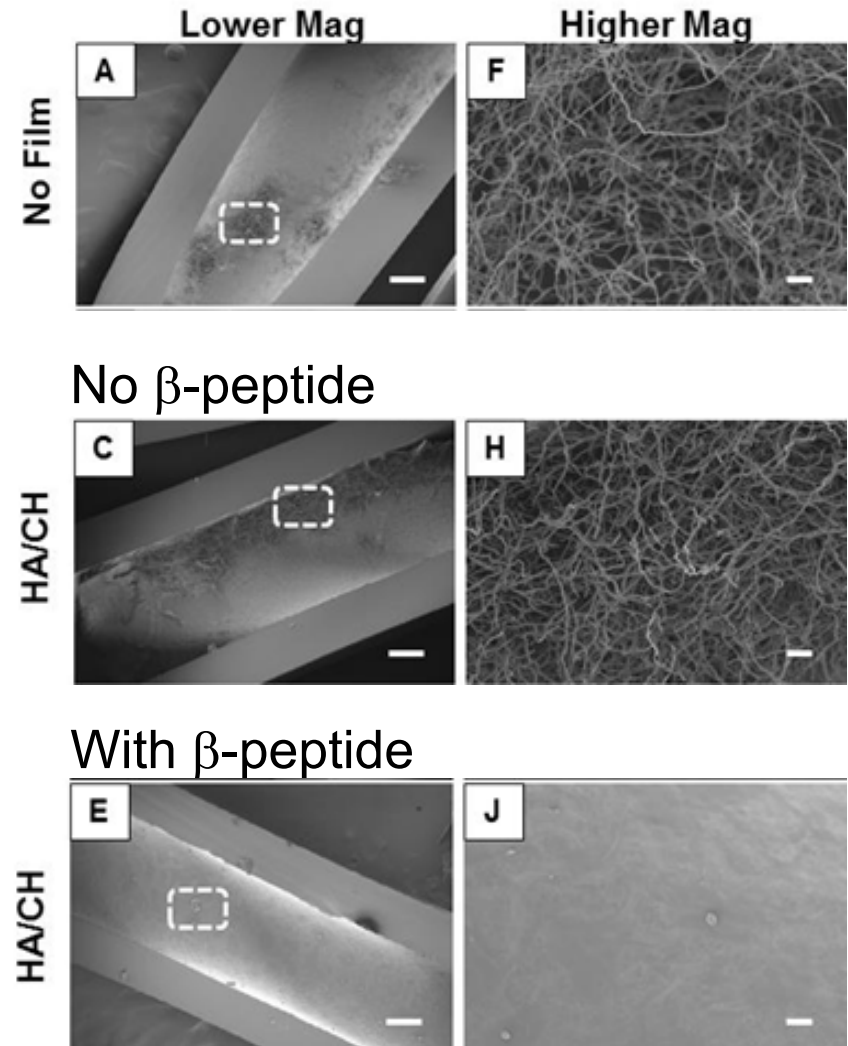
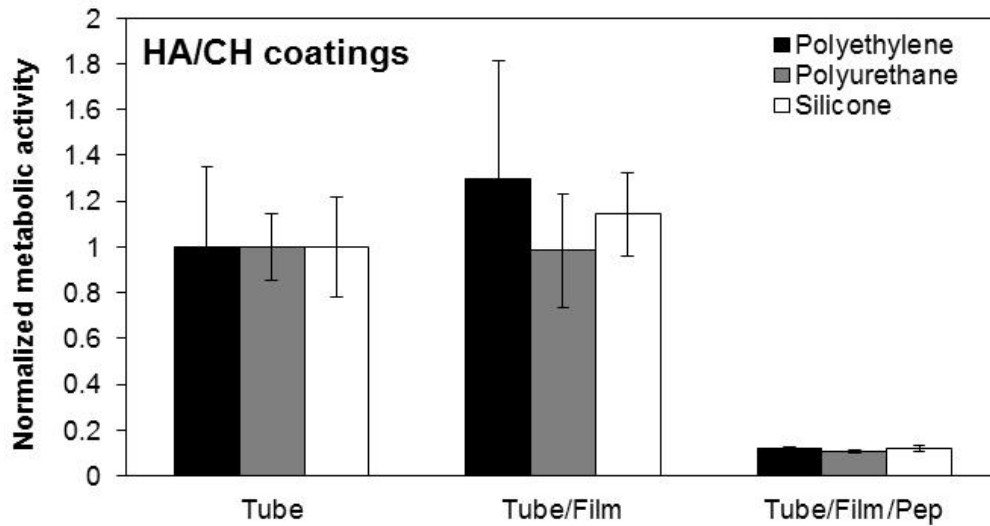
HA/CH

Fluorescent β -peptide



β -peptide release over 2-4 months

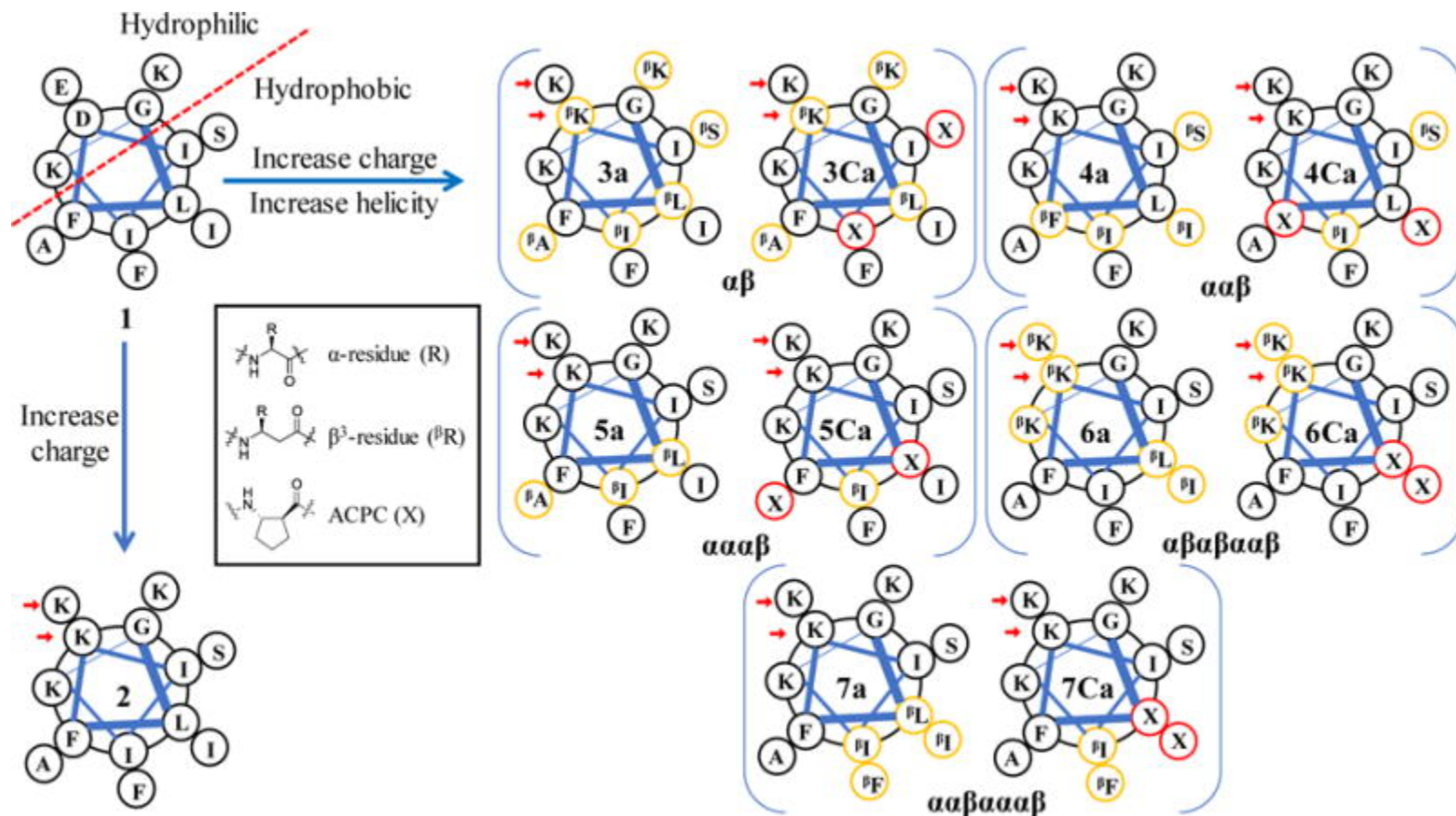
β -peptide-loaded catheters resist *C. albicans* biofilm formation in SU



HA/CH films loaded with β -peptide prevent *C. albicans* biofilm formation on catheters *in vitro*

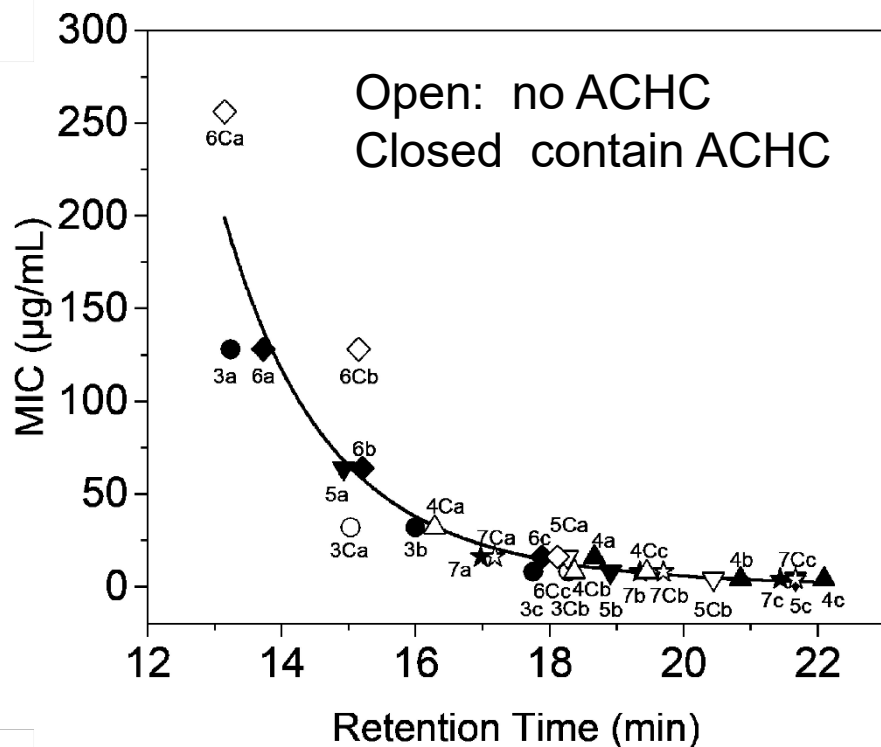
α/β -peptide analogues of aurein 1.2

α/β peptides mimic sequence and structure of aurein 1.2

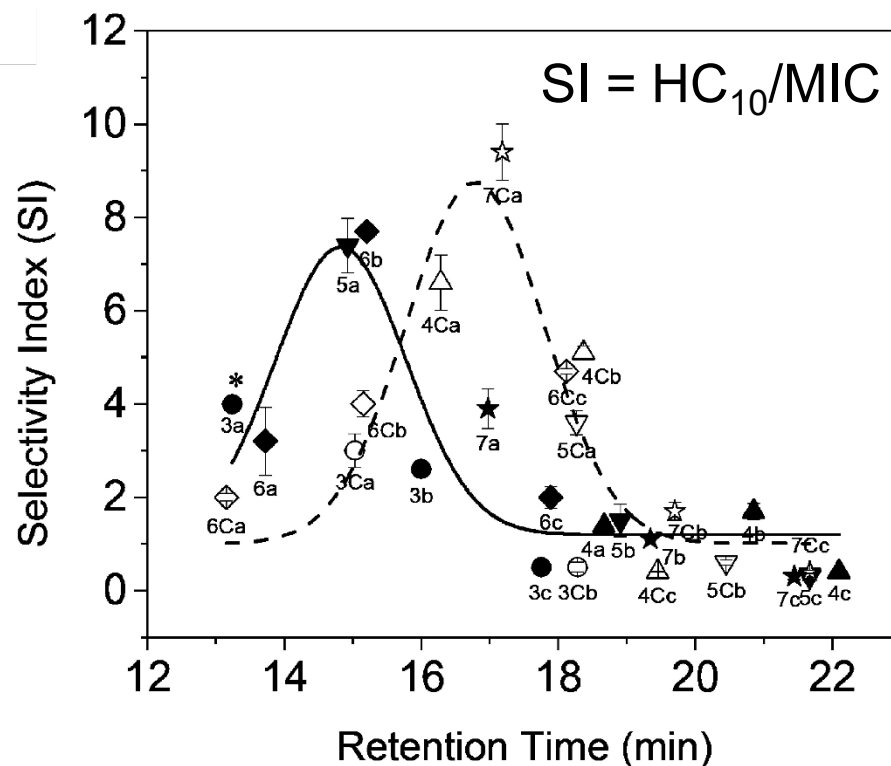


Active and selective α/β -aurein analogues

Peptide activity



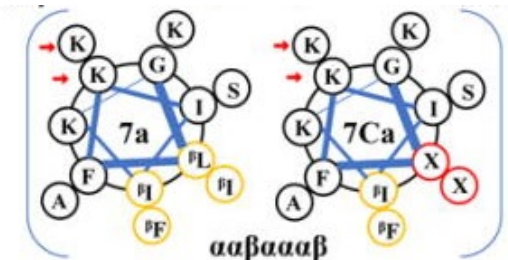
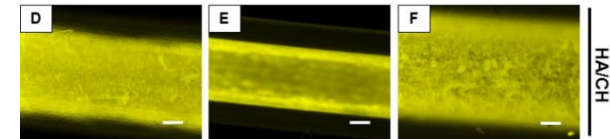
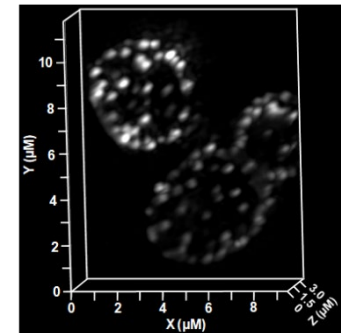
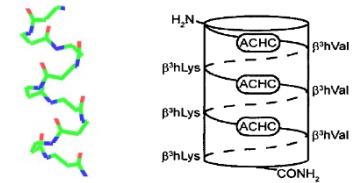
Peptide selectivity



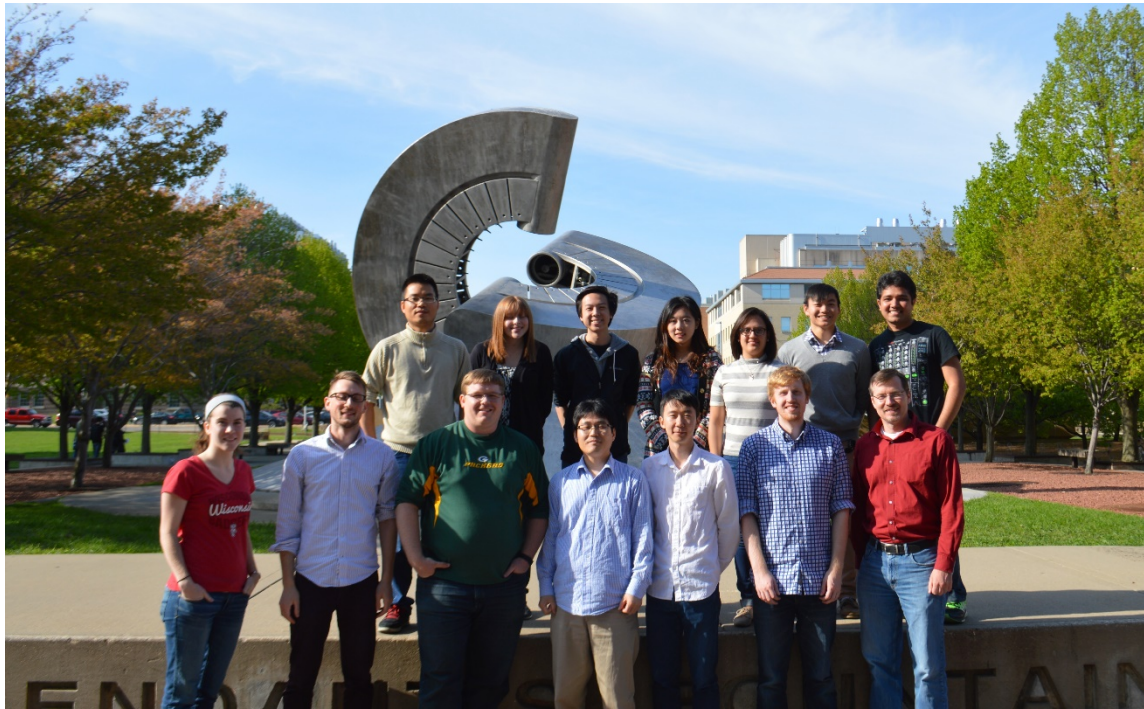
Active and selective α/β -aurein analogues at intermediate hydrophobicity

Summary

- Amphiphilic, helical β -peptides exhibit selective antifungal activity
- β -peptides kill *C. albicans* by plasma and intracellular membrane lysis
- Delivery of antifungal β -peptides from polyelectrolyte multilayers prevents *C. albicans* biofilms on catheters *in vitro* and *in vivo*
- α/β -analogues of aurein 1.2 have been engineered for selective antifungal activity



Acknowledgements



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Gellman



**Recruiting postdocs:
peptide engineering,
delivery, applications**